

Population density estimation of meso-mammal carnivores using camera traps without the individual recognition in Maduru Oya National Park, Sri Lanka

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Abstract

Reliable population estimates are crucial for the conservation and management of faunal species. Population data of meso-mammal carnivores in Sri Lanka, as well as elsewhere in the world, is scarce. We estimated population densities of meso-mammal carnivores in Maduru Oya National Park (MONP) using Random Encounter Model (REM) and Camera Trap Distance Sampling (CTDS) methods in this study. A total of 3,402 camera trapping days yielded 3,357 video captures of 69 different animal taxa including 658 video captures of meso-mammal carnivores. In this study, we recorded all 12 meso-mammal carnivore species found on the island. The two density estimate methods generated similar population estimates indicating that both methods are compatible to be applied in tropical forest habitats for meso-carnivore species. We identify MONP as an area with high richness for the focal species. The study also generated movement speed, activity patterns, activity levels, and day ranges for the focal species, which will be useful for future research. We discuss the population density estimates for different meso-carnivore species and the use of REM and CTDS density estimation methods and their applicability to a tropical meso-carnivore community.

Keywords: *Random Encounter Model, Camera Trap Distance Sampling, population monitoring, activity level, day range, species abundance*

Introduction

Accurate and updated population density estimates are vital for the proper evaluation of the conservation status of species, as well as for the management and decision-making about wildlife populations (Luo et al. 2020; Romairone et al. 2018; Jiménez et al. 2017; Royle et al. 2013; Carbone et al. 2006). Focused research on estimating mammalian carnivore populations remains scarce in Sri Lanka. Although there have been efforts on estimation of the population density of the Sri Lankan leopard (*Panthera pardus kotiya*) – the apex predator of the country (Webb et al. 2020; Kittle and Watson 2018; Kittle et al. 2017) – the population densities of many other species of mammalian carnivores have not been assessed (Kittle and Watson 2018; Miththapala 2018; Wijesinghe 2006; Weerakoon and Goonatilake 2006). In this study, we focused our work on estimating the population densities of meso-mammals of the order Carnivora (meso-carnivores/small carnivores) that inhabit Maduru Oya National Park in the dry zone of Sri Lanka. Meso-mammals are defined as “medium sized

47 mammals larger than rodents, up to roughly fox/jackal sized” (Parker et al. 2012; Hoffmann
48 et al. 2010), “which are between 150g-10kg in weight” (Morrison 2013).

49 Several factors such as the difficulty of individual recognition (Johansson et al. 2020) for
50 spatial capture recapture (SCR) density estimate models, nocturnal/elusive behaviour, solitary
51 activity and high costs of live-trapping methods (Hardouin et al. 2021; Romairone et al. 2018;
52 Sheftel 2018; O'Brien 2011; Rowcliffe et al. 2008; Silveira et al. 2003) have influenced the
53 lack of information for these species. Meso-mammals of the order Carnivora include an
54 ecologically important guild of species that plays key roles as predators, seed dispersers, as
55 well as influencers of community structures in tropical forest ecosystems, regulating lower
56 trophic levels and maintaining biodiversity (Hardouin et al. 2021; Kalle 2013; Kalle et al.
57 2013; Roemer et al. 2009). They are also considered carriers of diseases, agricultural pests
58 and apex predators in some ecosystems (Roemer et al. 2009). This group of mammals is
59 represented in Sri Lanka by the families Felidae (small wild cats), Herpestidae (mongooses),
60 Viverridae (civets), Mustelidae (otter) and Canidae (jackal). Within these families, there are
61 12 species (Annex I) in Sri Lanka (Hunter 2019; MoMD&E 2019; Dittus 2017; Weerakoon
62 2012).

63 With advancements in camera trapping technology, there has been a rise in research based on
64 camera trapping methods (Green et al. 2020; Meek et al. 2020; Glover-Kapfer et al. 2019;
65 Meek et al. 2014; O'Brien 2011). The scope of these studies spreads across a wide range of
66 different ecological facets such as faunal checklists, abundance, density estimations,
67 population monitoring, behavioural studies, species specific focal research studies and
68 wildlife management (Cappelle et al. 2021, Rovero et al. 2013; Meek et al. 2012; Bater et al.
69 2011; Zimmermann et al. 2011; O'Brien 2011; TEAM Network 2011; Cleverger et al. 2009;
70 Tobler et al. 2009; Bowkett et al. 2008; Rovero and De Luca 2007; Karanth et al. 2006;
71 Sanderson and Trolle 2005). However, there remained the absence of a reliable and cost-
72 effective method of population density estimation of mammalian fauna that cannot be
73 recognised individually (Chatterjee et al. 2020; Gilbert et al. 2020; Rowcliffe et al. 2008;
74 Srbeek-Araujo and Chiarello 2005). This lacuna was filled by the Random Encounter Model
75 (REM) developed by Rowcliffe et al. (2008) after the early efforts of occupancy-based
76 models (Royle and Nichols 2003) and *N*-mixture models (Royle 2004) for abundance
77 estimation. Since then, there has been several research studies that have been conducted
78 based on REM model (Palencia et al. 2021b; Pfeffer et al. 2017; Rademaker et al. 2016;
79 Manzo et al. 2012) as well as modified methods such as the Random Encounter and Staying

81 Time (REST) by Nakashima et al. (2017). Spatial count (SC) models (Chandler and Royle
82 2013), time-lapse based models (Moeller et al. 2018), spatial presence-absence (SPA) models
83 (Chatterjee et al. 2020; Ramsey 2015) and species space use (SPU) models (Luo et al. 2020)
84 for populations without markings are several other methods that were recently developed
85 each with their own or common limitations. With the rapid technological development of
86 digital camera traps, the video recording capability of camera traps and multiple snapshots
87 with faster trigger speeds have paved the way for development of REST model (Nakashima
88 et al. 2017) and recently, the modified camera trap distance sampling (CTDS) method (Howe
89 et al. 2017) of the well-known ‘Distance Sampling’ (DS) approach (Thomas et al. 2010;
90 Buckland et al. 2015, 2004, 2001).

91 Instead of using the auxiliary data such as day range determined by telemetry methods to
92 support the REM, during the last decade, this method has evolved to be self-supplemented
93 based solely on camera trapping information (Hofmeester et al. 2017; Rowcliffe et al. 2016,
94 2011, 2008). The process of calculating the species densities using REM generates several
95 important parameters such as animal speed, activity level and day range, which then supports
96 a variety of ecological studies. Therefore, REM has provided a means to investigate a wider
97 range of ecological parameters to assist in the species conservation and management.

98 After the modifications of Howe et al. (2017), the DS method – which has been well
99 established over the years – can also be used to determine species densities even when
100 individual markings are absent. Distance sampling can be considered one of the most applied
101 methods for monitoring of wildlife populations (Buckland et al. 2015; Buckland et al. 2001;
102 Thomas et al. 2010). However, the traditional DS method was more applicable for species
103 that could be detected easily and directly during the surveys (Corlatti et al. 2020; Buckland et
104 al. 2015). When it comes to rare, elusive and smaller animal species, the applicability was
105 low (Corlatti et al. 2020; Marques et al. 2013). As a result, in the recent past, there has been a
106 rise in usage of passive DS methods such as sonar, radar and acoustic surveys (Corlatti et al.
107 2020; Buckland et al. 2015; Marques et al. 2013). The implementation of CTDS (another
108 passive DS method) can be considered a revolution in the wildlife population monitoring
109 study methods, as it greatly reduces the limitations that previously prevailed. Availability of
110 user-friendly software and R packages together with adequate methodologies and literature
111 will make CTDS more popular in future camera trap based research work. Since its
112 introduction, CTDS method has generated reliable density estimates in most of the recent

114 studies (Cappelle et al. 2021; Palencia et al. 2021b; Bessone et al. 2020; Harris et al. 2020;
115 Cappelle et al. 2019).

116 In this study, the SCR methods where individual recognition is required were not selected,
117 because there were no identifiable pelage patterns in most of the focal species except for the
118 Felids. Therefore, as the best alternatives, we selected REM and CTDS methods of density
119 estimation using camera traps. Most of the recent REM and CTDS camera trapping
120 applications have focused on larger ungulate species (Pal et al. 2021; Pfeffer et al. 2017;
121 Rovero and Marshall 2009) or on single species (Corlatti et al. 2020; Harris et al. 2020;
122 Cappelle et al. 2019; Gray 2018; Cusack et al. 2015; Anile et al. 2014; Engeman et al. 2013;
123 Manzo et al. 2012). Rich et al. (2019) investigated population density of multiple forest
124 carnivore species, using SCR methods. The number of camera trap studies on population
125 densities of meso-mammal carnivores remains low and CTDS based multi-species
126 evaluations of this group of fauna are limited (Cappelle et al. 2021; Palencia et al. 2021b;
127 Hardouin et al. 2021; Bessone et al. 2020). Therefore, this is one of the early applications of
128 these new methods to a tropical meso-carnivore community and the first multi-species
129 density estimation in Sri Lanka.

130 The objectives of this study were; i) to generate density estimates for the meso-mammal
131 carnivores in MONP; ii) to compare the density estimates derived from REM and CTDS
132 methods and assess their applicability in practical situations. During the process of generating
133 density estimates, we developed activity levels, activity patterns, day range, and detection
134 radius/distance parameters for the focal species. Hence, the results generated through this
135 study will provide a range of information to fill research gaps and to benefit future
136 conservation and management requirements.

137 **Materials and Methods**

138 **Study area**

139 We conducted this study in Maduru Oya National Park (588 km²) situated in the dry zone
140 (predominantly, in the northern and eastern parts of the country) (Punyawardena 2020) of Sri
141 Lanka. We carried out camera trapping in the western flank of the park adjacent to the
142 western bank of the Maduru Oya reservoir situated in the centre of the park (Fig. 1). The area
143 of study was 304km² – comprising grasslands, shrublands and the climax habitat of dry mixed
144 evergreen forest. Rocky outcrops can be observed in patches scattered throughout the park

(Jayasekara et al. 2021). Most of the grasslands and shrublands are a result of slash and burn cultivation practised over the years, until the area was declared a national park in 1983 (IUCN 1990). The grasslands assume characteristics of savannas in some areas, whereas the reservoir perimeter is surrounded by seasonal grasses that grow during the dry season (late January-October). The park is well known for large numbers of sightings of Asian elephants (*Elephas maximus*) and also provides habitats for many other mammalian species (Jayasekara and Mahaulpatha 2019) as well as avifauna (Dissanayake 1995). The large Maduru Oya reservoir (6,100ha), constructed as a part of the Mahaweli Development Project (a large-scale national irrigation project to harness water from Sri Lanka's largest river – the Mahaweli), situated at the centre of the park, has a considerable influence on this faunal assemblage and creates a large perimeter (97.8 km) with aquatic, riparian habitats. We selected the western flank of the park for our study because the natural barriers and the man-made reservoirs/canals help in fulfilling one key assumption of both REM and CTDS models – the requirement of a closed population (Howe et al. 2017; Rowcliffe et al. 2008). Most of the study area is surrounded by four large reservoirs, irrigation canals, rock formations, and cultivated lands surround (Fig. 1) (IUCN 1990).

Camera trapping

We conducted camera trapping mostly during the dry season (compared to the monsoon season from October to January) (IUCN 1990) adhering to the protocol for tropical forest vertebrate camera trap survey by Team Network (2011). We divided the selected study area into 2 x 2 km plots using a feature grid in ArcMap version 10.4.1 (Esri, Redlands, USA) (Fig. 1). Generating this grid fulfils the spacing requirement recommended by Team Network (2011) of placing one camera in every 2 km² grid plot. We used two infra-red-triggered camera models: Browning Strike Force HD Pro (n=10, low glow flash) and Browning Dark OPS HD Pro (n=15, no glow flash) (Browning, USA). Except for the type of flash, the specifications of the two camera models were similar. We especially used these flash types to reduce interference to animals and meet the assumption of independent animal movement (Rowcliffe et al. 2008).

We established camera trap stations in 90 plots. We excluded plots covered with large areas of reservoir, inaccessible terrain and some plots with repetitive habitats, to obtain a balanced sampling effort in all available habitat types (Rovero et al. 2013). We deployed the moving survey method (Palencia et al. 2021b) to better use the available cameras which increase the

179 sampling effort and precision. One station had to be excluded from analyses because a
180 camera was stolen by a poacher, reducing the total sampling points to 89. We randomly
181 selected plots and we placed cameras within each selected plot moving in a random distance
182 from a random starting point in the grid line of the plot grid (walking perpendicularly to the
183 grid line). This randomisation of camera stations fulfils the requirement of both REM and
184 CTDS methods. Usually, we attached cameras at a fixed height of 25 cm to tree trunks or an
185 erected log. We selected this height based on previous literature (Kalle 2013) and our field
186 experience of camera trapping meso-mammal carnivores, to maximise detection. We oriented
187 cameras in a northward direction. We had to deviate the realised sampling locations and
188 orientations up to a maximum of 100 m and 40° respectively to ensure cameras were
189 mounted at suitable locations without obstructions (Pfeffer et al. 2017; Howe et al. 2017).
190 However, we remained as close as possible to the predefined coordinates and orientation. We
191 ensured mounting cameras parallel to the ground and to avoid areas with slopes, to obtain
192 accurate distance measurements during analyses. We used protective metal cases and python
193 lock cables when mounting cameras, to reduce damage from elephant attacks and theft. We
194 set all cameras to function for 24 hrs in a stretch of 38.2 days on average. We set the range
195 parameter to “long range”, mode of capture to “video” and trigger delay to one second. These
196 specifications ensured that capture data could be used for both the REM and CTDS methods.
197 We monitored the camera stations on a routine basis of 10-15 days and stations with defects
198 in cameras/memory cards were resampled to obtain the desired sampling effort. We had to
199 reassign two camera stations where initial coordinates coincided with resting places of a
200 fishing cat and ring-tailed civets.

201 **Random Encounter Model**

202 We used REM developed by Rowcliffe et al. (2008) as one method of meso-mammal
203 carnivore density ($D \text{ km}^{-2}$) estimation. The equation,

$$204 \quad D = \frac{y}{t} \times \frac{\pi}{v * r * (2 + \theta)}$$

205 is used for the calculation where y denotes the number of capture events; t , the survey effort
206 (camera trapping days); v , the average daily distance travelled (km/day); r , the average
207 distance to the first capture of animals (km); and the average angle to the capture animals is
208 θ (radians). The daily distance travelled (v , day range) is derived using the movement speed
209 (s) and activity level (a) of animals following the equation shown below.

$$v = s \times a$$

The movement speed (s) of each animal was derived using the simple equation $s_i = \frac{d_i}{t_i}$ (Pfeffer et al. 2017) where d_i denotes the distance travelled and t_i the time duration. We followed the procedure described by Rowcliffe et al. (2016) to calculate the average speed parameter by fitting probability distributions to samples of individual speed observations obtained from video captures instead of multiple snapshots. The R package ‘*fitdistrplus*’ (Delignette-Muller and Dutang 2015) was used for model fitting and best fitting models were selected based on Akaike Information Criterion (AIC) values.

To determine activity level (a) and the proportion of the day a species is active (Rowcliffe et al. 2016), we used the R package ‘*activity*’ (Rowcliffe, 2019; Rowcliffe et al. 2014). We converted the time stamp data of species captured on camera trap videos to radian time and analysed this in R with 1,000 iterations.

To determine the radial distance (r) to the capture animal and d_i , accurate evaluation of distance from the camera was highly important. The method generally used for distance estimation is based on marking certain distance intervals from the camera at the time of mounting camera traps (Palencia et al. 2021b; Pfeffer et al. 2017; Caravaggi et al. 2016) or measuring distances of each animal manually at time of dismounting (Rowcliffe et al. 2011). However, we found that this method required extra time and effort in the field and that visual estimation of distances outside the marking points was difficult. In addition, in MONP where elephant activity was quite high, spending extended time in certain locations was dangerous. Therefore, we deviated from the original method of measuring distance. Rather than measuring distances on location, we incorporated the distance intervals in a pre-marked grid (Caravaggi et al. 2016) (Fig. 2), as a standard which could be superimposed on all camera trap records. This method made the determination of distances and trigonometric calculation of distances (Pfeffer et al. 2017; Caravaggi et al. 2016) easier and accurate (a distance-angle table generated following this method is given in Annex II. We calculated the time difference (t_i) from the time difference recorded in each video capture. Instead of camera specific detection distance and angles (Rowcliffe et al. 2008), for our analyses, we used species specific average detection distances (ADD); average detection angles (θ , ADA) derived exclusively from camera trap captures (Pfeffer et al. 2017). Because most of the observed species are solitary species we did not apply the group size function to the density equation

(maximum average group size recorded was 1.06). We performed density calculations in R version 4.0.3 (R Core Team 2013) bootstrapping with 1000 iterations from the original data.

Camera trap distance sampling method

The CTDS method, developed by Howe et al. (2017), follows the standard point transect methods (Buckland et al. 2001) and each camera station is considered a sample point. Density (D) is estimated as,

$$\hat{D} = \frac{\sum_{k=1}^K n_k}{\pi w^2 \sum_{k=1}^K e_k \hat{p}_k}$$

where, k = the camera station/point

K = set of camera stations/points

n = number of captures

w = truncation distance beyond which any recorded distances are discarded

e_k = effort expended at point k

\hat{p}_k = estimated probability of obtaining an image of an animal that is within θ and w in front of the camera at a snapshot moment

The effort is described by $e_k = \frac{\theta T_k}{2\pi t}$, multiplied by the activity level (a), yields the actual trapping effort as follows.

$$e_k = \frac{\theta T_k}{2\pi t} * a$$

θ = average detection angle

T_k = time period the camera was active

t = the time between two snapshot moments considered (within the video)

Detailed explanations of these equations are provided in Howe et al. (2017). We calculated the effort for each camera trap station for separate species and provided it as the input for effort in distance software. Because of low height of the camera mount, the w values exceeding 6.2m were less accurate. Therefore, we right truncated to a maximum of 6.2m and

268 left truncated to 1m. We used the previously calculated a values (for REM) in this equation.
269 Detection angle θ was estimated as 0.715585 radians. We recorded the distance between
270 cameras and animals every three seconds in video captures, 24 h per day. Hence, parameter t
271 applied in the above equation was three seconds. A special consideration was given for
272 observations of reactivity to the cameras by the animals. In such cases, the latter part of the
273 videos where animals unusually stayed extended time periods in front of the cameras were
274 excluded from analysis. We used the “Distance 7.1” software package (Thomas et al. 2010)
275 for density calculations. Half-normal and hazard rate candidate models of the detection
276 function were tested setting the maximum adjustment parameter at one to reduce overfitting
277 with overly complex models (Cappelle et al. 2021; Howe et al. 2019; Buckland et al. 2010,
278 Marques et al. 2007). Fitted probability density and detection probability plots were inspected
279 to ensure they were monotonically non-increasing (Cappelle et al. 2021, Howe et al. 2019).
280 Competing models with sufficient goodness of fit were selected using AIC criteria.

281 We estimated variances in Distance 7.1 using the default analytic variance estimators based
282 on detection probability and encounter rate (Fewster et al. 2009), and also from 1000 non-
283 parametric bootstrap resamples of camera station data points (Cappelle et al. 2021; Howe et
284 al. 2017; Buckland et al. 2001). Bootstrap density estimates were recorded separately.
285 Coefficient of variation (CV) was obtained using the square root of the variance and the point
286 estimates in all methods used.

287 Density estimations were compared statistically using the Wald test, with a test statistic W
288 assessed on the chi-squared distribution with one degree of freedom (Palencia et al. 2021b;
289 Wald and Wolfowitz 1940).

290 We calculated relative abundance index (RAI) as a crude estimate (Cappelle et al. 2021) for
291 all species, especially to represent the less abundant species where sufficient samples were
292 lacking to calculate density. RAI was calculated as encounters per hundred trap nights (Kalle
293 2013).

294 **Results**

295 **Meso-carnivore assemblage and capture abundance**

296 A total of 3,402 camera trapping days yielded 3,357 video captures of 69 different animal
297 taxa including 658 video captures of meso-mammal carnivores. During this study in MONP,

we recorded all 12 meso-mammal carnivore species (Annex I) found on the island. However, we captured only seven species in excess of 45 videos. The abundance of rusty spotted cats ($n=4$) (Fig. 3 A), jungle cats ($n=5$), common palm civets ($n=2$), and brown mongooses ($n=10$) was very low in the study site. Therefore, density calculations based on REM model were performed only for the remaining species: fishing cats ($n=106$) (Fig. 3 C), ruddy mongooses ($n=302$), stripe-necked mongooses ($n=52$) (Fig. 3 B), ring-tailed civets ($n=118$) (Fig. 3 D), golden palm civets ($n=45$) (Fig. 3 E), otters ($n=46$) and golden jackals ($n=45$). We estimated that these capture numbers are greater than, or closer, to the benchmark of ‘around 50’ captures recommended by Rovero et al. (2013) for REM density estimates. Density calculations for the same species were also conducted based on CTDS method.

Detection distances/movement speeds/day ranges, activity patterns and activity level

The average detection distance (ADD) value ranged from 1.90 – 4.07 m for the species considered. The rusty-spotted cat recorded the lowest distance value, while otter recorded the highest. In general, effective detection distance (EDD) values were greater than the observed ADD values except for golden palm civet (Table 1).

The movement speeds ranged from 0.72 - 3.42 km/h. The fastest moving species was the otter, followed by the golden jackal, resulting in high day ranges for those two species. The highest activity levels were shown by fishing cat and golden jackal indicating that they were active during a greater proportion of time when compared to other species. We observed that all mongoose species and golden jackals were diurnal while civet species and otters were nocturnal (Table 1; Fig. 4). Fishing cats were mostly nocturnal yet could also be observed during day time as well. Jungle cats and rusty-spotted cats were recorded mostly at night. The highly nocturnal golden palm civet was the least active species. In addition, we observed this species to be the second slowest, recording the lowest day range (3.47 km/day).

Comparison of REM and CTDS density estimates

Based on Wald test statistic, any of the density estimates obtained from different methods of analyses were not significantly different for any of the species ($p>0.05$). However, the density estimates of fishing cat (Wald test: CTDS vs. REM: $W = 0.91$, $p = 0.34$) and ring-tailed civet (Wald test: CTDS (b) vs. REM: $W = 2.06$, $p = 0.15$) obtained using REM were relatively higher than CTDS estimates (Table 2). Ruddy mongooses had the highest abundance, and it was among the highest density estimates in all three analyses. However, the

331 REM density estimate of ring-tailed civet was the highest recorded density. Lowest densities
332 were recorded for otter and golden jackal. Density estimates derived using the CTDS method
333 generally yielded lower figures when compared to the REM method (except on two occasions)
334 (Table 2). However, the coefficient of variation (CV) values were generally higher in the
335 CTDS method compared to REM (except in one occasion) (Table 2). The low abundance of
336 rusty-spotted cat, jungle cat and brown mongoose were indicated by very low RAI values
337 (Table 2).

338 Discussion

339 Our findings show that MONP is a protected area with a rich assemblage of meso-mammal
340 carnivores (Annex I). However, when Felid species were considered, there were very few
341 jungle cat and rusty-spotted cat camera trap sightings inside the study area of MONP.
342 Because of the low number of captures of those two species, we were unable to calculate
343 population densities using the REM or CTDS. However, RAI values of jungle cat and rusty-
344 spotted cat were the lowest among the species on which we focused. The limited number of
345 records of rusty spotted cats and jungle cats were from dense dry mixed evergreen forests and
346 shrublands respectively, conforming the findings of Bora et al. (2020), Chatterjee et al. (2020)
347 and Palei et al. (2019) on these cats' habitat occupancy. Based on our field observations, we
348 posit tentatively that one reason for the low abundance could be that these two species are
349 attracted to agricultural areas (paddy fields) and habitat edges, alternative habitats with
350 abundant small mammal prey used by both species (Dharmarathne, personal communication,
351 2021; SCAR 2021; Bora et al. 2020; Miththapala 2018; Šálek et al. 2010; Nekaris 2003). In
352 contrast, our results indicate that MONP is home to a healthy population of fishing cats, the
353 largest of the three felid species studied. The fishing cat population densities recorded in this
354 study are among the highest densities recorded for the species compared to research in other
355 countries (Mishra et al. 2018; Sathiyaselvam et al. 2016). The large Maduru Oya reservoir
356 and other reservoirs within the park provide ample food for this carnivore that is associated
357 with water (SCAR 2021; Ganguly and Adhya 2020; Hunter 2019; Miththapala 2018;
358 Mukherjee et al. 2016). The frequent release of fingerlings to the Maduru Oya reservoir by
359 the local community-based fishing society and the abundance of fish and aquatic avifauna in
360 its habitats make MONP an ideal site for fishing cats through the provision of food resources
361 (Ganguly and Adhya 2020; Hunter 2019; Cutter 2015; Kitchener et al. 2010; Haque and
362 Vijayan 1993).

364 Our results show that density of otters was relatively low, although this is another species that
365 prefers aquatic fauna as its main prey (Dettori 2021; Romero and Guitián 2017; Bouros and
366 Murariu 2017; de Silva 1996; Carss 1995). Although their population density (maximum
367 estimate 0.16 per km²) is similar to estimations from other studies (Quaglietta et al. 2015;
368 Hájková et al. 2009; Lanszki et al. 2008), there may be a foraging niche overlap with fishing
369 cat given their known food habits (Dettori 2021; Ganguly and Adhya 2020; Hunter 2019;
370 Cutter 2015; de Silva 1996; Kitchener et al. 2010; Carss 1995; Haque and Vijayan 1993).

371 This is likely the first effort of estimating the densities of civets and mongooses in a wild
372 habitat in Sri Lanka. The grey mongoose, which is thought to be common in the northern
373 third of the island (Wijeyeratne 2008), was not captured in camera traps, although through
374 direct visual observations, we spotted a couple of individuals. Santiapillai et al. (2000) and
375 Wijeyeratne (2008) have reported a similar situation from Yala National Park, which is
376 another protected area situated in the dry zone of the country. The brown mongoose
377 abundance in MONP was low. The density of stripe-necked mongoose was moderate. We
378 obtained high population density estimates among all focal species for the ruddy mongoose,
379 which was also the dominant mongoose species in MONP, as observed by Jayasekara and
380 Mahaulpatha (2019). The ring-tailed civet was the Viverrid with the highest density,
381 validating its least concern (LC) status in the National Red List (MOE 2012). The common
382 palm civet density was not calculated because of the very low number of captures.

383 When the two main analysis methods (REM and CTDS models) are compared, the only
384 contrasting result we obtained was the density of endemic golden palm civet. Golden palm
385 civets are generally arboreal (Wijeyeratne 2008) and the camera traps capture them only
386 when they are on the ground. Therefore, the speed estimation based on a 2D model becomes
387 biased, because their vertical movements were not recorded through our camera arrangement.
388 The slowness of golden palm civets on ground is indicated by our speed calculation of 0.89
389 km/h. Considering the above, we recommend that the CTDS estimates (0.80-0.97 individuals
390 per km²) in which speed is not a parameter, to be relatively more accurate for this species
391 despite the drawback of not recording arboreal movements. However, the bias caused by not
392 recording vertical movements would not be completely eliminated unless methodology is
393 adapted to account for such complex scenarios. In general the CTDS method is considered
394 more suitable for low abundant species (Palencia et al. 2021b).

396 The unusually high “speed parameters” generated for otter and golden jackal did not have an
397 adverse impact on REM density estimation because we obtained similar densities from the
398 CTDS method. We suggest estimating the day ranges of the above two species in the study
399 area using another method/repeated method to confirm the values we received. However,
400 according to Rowcliffe et al. (2012) and Palencia et al. (2019) the alternative methods such as
401 telemetry often underestimate travel distances. Radio tracking studies of otter in other
402 countries indicate that otters can cover long distances ranging from >20 – 100 km in a single
403 day (Ruiz-Olmo 2001) and occupy large home ranges (Quaglietta et al. 2015). Therefore, the
404 day range of 27.5 km observed in the present study could likely be accurate. Research
405 focused on golden jackal in Sri Lanka remains scarce (Jayaratne and Seneviratne 2020) and
406 the observed density value was within the density range observed by Šálek et al. (2014) in
407 Balkan Peninsula.

408 Approximately similar density estimates generated by both analyses, despite REM estimates
409 being slightly higher, conform the observations of Palencia et al. (2021b). Therefore, we
410 recommend both REM and CTDS methods for the population density estimation for meso-
411 mammal carnivores in tropical habitats. However, CV values of CTDS method were
412 relatively higher than the REM values despite the similarities of density figures. Density
413 estimates of species with CV values <40% are generally considered reasonable, and in recent
414 research work, the effort has been to further increase the precision (Cappelle et al. 2021;
415 Palencia et al. 2021b; Harris et al. 2020; Howe et al. 2019). According to Cappelle et al.
416 (2021) CV values between 10-20% are more desirable. When the present study is considered,
417 61.9% of the CV values were <40% and 42.9% were <30%. According to recent research, the
418 precision can be further increased by increasing the sampling effort in different ways
419 (Cappelle et al. 2021; Rovero et al. 2013). Hence, in order to obtain a greater number of
420 capture events, we suggest following the recommendations of Cappelle et al. (2021); (a)
421 increase the number of camera stations or (b) increase the length of sampling period.
422 However, there remains the logistic concerns that are associated when camera trapping
423 extremely rare species. We suggest the length of sampling period to be increased while
424 deploying the appropriate number of camera stations as the best way forward. The moving
425 survey method we followed also reduced the limitation occurred by low number of cameras,
426 increasing the effort and precision.

427 Accounting for overdispersion with more customized model selection criteria as described by
428 Howe et al. (2019) would increase the accuracy and precision of CTDS results. We identified
429

430 that proper estimation of movement speed, activity and ultimately the day range of species
431 was critical for the final density results of REM. Application of recently developed method
432 by Palencia et al. (2019) integrating the behaviors and speed-ratio in calculations makes it
433 possible to obtain unbiased day range values. Furthermore, with the development of machine
434 learning techniques (Palencia et al. 2021a) and specialised “R packages” like
435 “trappingmotion” (Palencia 2020) the analysis process will be streamlined. However, dealing
436 with multiple species, we observed that number of encounters need to be higher in order to
437 apply this method. When monitoring gregarious species, it is recommended to consider
438 applying the group size function in the density equations (Rowcliffe et al. 2008). During the
439 present study, the ruddy mongoose and the golden jackal were the species with the highest
440 average group size with a value closer to one (1.06). Therefore, we did not include group size
441 in the analyses.

442 We used a modified distance measuring method for this study, which saved the time and
443 effort during field work and further helped to obtain accurate measurements during analyses.
444 However, we would like to highlight that if the distance grid and table are used, camera
445 height and orientation should be positioned precisely. In addition, based on the camera
446 mounting height, this distance grid and table can be generated easily prior to camera trap
447 deployment in the field. It is also important to note that the focal distance of the camera may
448 differ from one model to another. When using different camera models, model specific
449 distance calculations should be used. This method is less applicable in complex field
450 situations with slope and rugged terrain. In those instances, original distance measuring
451 techniques or slope adjusted parameters can be used. Both REM and CTDS methods require
452 reasonable amount of field effort as well as substantial amount of time for processing the
453 images/videos and exploratory analyses (Palencia et al. 2021b). We would like to highlight
454 the requirement of suitable software for image and especially video processing. Integration of
455 such software with machine learning would greatly reduce the time required in computer
456 analyses.

457 The type of camera flash also has an impact on the behaviour and the movement speed of the
458 animals. We highly recommend a no glow flash model such as Browning Dark OPS HD Pro,
459 which causes minimum interference to the animals when REM and CTDS methods are used.
460 However, we observed that the low glow flash Browning Strike Force HD Pro also interfered
461 less, except for a few observations which we had to discard the capture records as
462 behavioural changes were observed. Selection of these flash types also increases the battery

464 life of cameras (one set of batteries usually lasted more than two months on video mode
465 during our study). We do not recommend white flash camera models. Most of the focal
466 species did not react to the cameras in a greater proportion of encounters. However, there
467 were several instances where fishing cats and ring-tailed civets were observing the cameras in
468 an enthusiastic nature where we had to discard some parts of the videos. Though not focused
469 on in this study, elephants were highly reactive to the cameras and were often found attacking
470 them. The use of videos (Cappelle et al. 2021; Howe et al. 2017) – instead of snapshots used
471 in early REM and CTDS based studies (Pfeffer et al. 2017; Rovero and Marshall 2009) –
472 improves the accurate identification of species. Moreover, the ability to observe the actual
473 behaviour of the animal helps to determine when reactive behaviours take place. This also
474 helped us to identify resting places of animals, which led to redeployment of two camera
475 stations. Because we assessed multiple meso-carnivore species in this study, there was the
476 concern of selecting a camera height that suits all species. Based on our observations, the
477 increase in species shoulder height did not adversely impact the detection or encounter rate.
478 Sometimes, the species could be identified even when some parts of the animal were out of
479 the frame (for example the Jackal). We selected the height of 25cm to reduce the bias caused
480 by not encountering the smaller animals when they are very close to the camera (for example
481 the rusty-spotted cat). Therefore, the selection of camera height should be based on the
482 morphometrics of the focal species. The availability of in-built display with video playback
483 option was very useful during routine observations in the field. In addition, with videos, the
484 movement speed estimation becomes more accurate because the bias caused by the delay
485 between the snapshots is removed. Even though the methods followed during our work would
486 have reduced the bias of animal reactivity and other technical concerns, we acknowledge that
487 they were not eliminated completely.

488 Our study provides population density estimates for the meso-mammal carnivore species in
489 MONP, which would inform future conservation and management decision-making and also
490 a template by which their status could be assessed in forest habitats in other parts of the
491 island. Additional parameters such as movement speed, activity patterns, activity level and
492 day range that we generated can be also used for future research in a broad range of
493 applications. The study shows clearly that REM and CTDS methods can be applied
494 practically under field conditions of tropical forests, to assess multiple species. The
495 recommendations for modifications to build upon original methodologies and analyses will
496 improve efficiency and cost-effectiveness of similar research in the future.

Conclusions

The study identifies MONP as a protected area with a rich meso-mammal assemblage. However, our study indicates that species such as rusty-spotted cat, jungle cat, brown mongoose, otter and golden jackal have low abundances and population densities. MONP sustains considerably healthy populations of fishing cats, ring-tailed civets and ruddy mongooses. The two main population estimation methods we used, the REM method and CTDS method could be applied successfully in the forest habitats of Maduru Oya. The CTDS method was more easily applicable in the field with suggested modifications of distance estimations. However, the relatively complex REM method can be more useful as it generates additional information such as activity, day range and movement speed which are useful for other ecological studies and decision making.

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Author contributions

524 Dharshani Mahaulpatha, Dulan Jayasekara and Sriyanie Miththapala conceived the study.
525 Dulan Jayasekara and Dharshani Mahaulpatha devised the methodology and established the
526 camera stations. Dulan Jayasekara analyzed the data with input from Dharshani Mahaulpatha
527 and Sriyanie Miththapala. Dulan Jayasekara led the writing of the manuscript. All authors
528 contributed critically to the drafts and gave final approval for publication.

529 **Competing interests**

530 Authors declare no competing interests

531 **References**

532 Anile, S., Ragni, B., Randi, E., Mattucci, F., Rovero, F., 2014. Wildcat population density on
533 the Etna volcano, Italy: a comparison of density estimation methods. *J. Zool.* 293(4):
534 252-261.

535 Bater, C.W., Coops, N.C., Wulder, M.A., Hilker, T., Nielsen, S.E., McDermid, G., Stenhouse,
536 G.B., 2011. Using digital time-lapse cameras to monitor species-specific understorey
537 and overstorey phenology in support of wildlife habitat assessment. *Environ. Monit.*
538 *Assess.* 180(1): 1-13.

539 Bessone, M., Köhl, H.S., Hohmann, G., Herbing, I., N'Goran, K.P., Asanzi, P., Da Costa,
540 P.B., Dérozier, V., Fotsing, E.D., Beka, B.I., Iyomi, M.D., 2020. Drawn out of the
541 shadows: Surveying secretive forest species with camera trap distance sampling. *J Appl*
542 *Ecol.* 57(5): 963-974.

543 Bora, J.K., Awasthi, N., Kumar, U., Goswami, S., Pradhan, A., Prasad, A., Laha, D.R.,
544 Shukla, R., Shukla, S.K., Qureshi, Q., Jhala, Y.V., 2020. Assessing the habitat use,
545 suitability and activity pattern of the Rusty-spotted Cat *Prionailurus rubiginosus* in
546 Kanha Tiger Reserve, India. *Mammalia* 84(5): 459-468.

548 Bouros, G., Murariu, D., 2017. Comparative diet analysis of the Eurasian otter (*Lutra lutra*)
549 in different habitats: Putna-Vrancea Natural Park and Lower Siret Valley, south-eastern
550 Romania. North West J Zool. 13(2): 311–319.

551 Bowkett, A.E., Rovero, F., Marshall, A.R., 2008. The use of camera-trap data to model
552 habitat use by antelope species in the Udzungwa Mountain forests, Tanzania. Afr J
553 Ecol. 46(4): 479-487.

554 Buckland, S.T., Anderson, D.R., Burnham, K.P., Laake, J.L., Borchers, D.L., Thomas, L.,
555 2001. Introduction to Distance Sampling. Estimating Abundance of Biological
556 Populations. Oxford University Press, Oxford, UK.

557 Buckland, S.T., Anderson, D.R., Burnham, K.P., Laake, J.L., Borchers, D.L., Thomas, L.,
558 2004. Advanced Distance Sampling. Estimating Abundance of Biological Populations,
559 OxfordUniversity Press, Oxford, UK.

560 Buckland, S.T., Rexstad, E.A., Marques, T.A., Oedekoven, C.S., 2015. Distance Sampling.
561 Methods and Applications, Springer, Heidelberg, Germany.

562 Cappelle, N., Després-Einspenner, M., Howe, E. J., Boesch, C., & Kühl, H.
563 S. (2019). Validating camera trap distance sampling for chimpanzees. Am. J.
564 Primatol. 81(3): e22962. <https://doi.org/10.1002/ajp.22962>

565 Cappelle, N., Howe, E.J., Boesch, C., Kühl, H.S., 2021. Estimating animal abundance and
566 effort–precision relationship with camera trap distance sampling. Ecosphere 12(1):
567 p.e03299.

568 Caravaggi, A., Zaccaroni, M., Riga, F., Schai-Braun, S.C., Dick, J.T., Montgomery, W.I.,
569 Reid, N., 2016. An invasive-native mammalian species replacement process captured
570 by camera trap survey random encounter models. Remote Sens. Ecol. Con. 2(1): 45-58.

- 572 Carbone, C., Christie, S., Conforti, K., Coulson, T., Franklin, N., Ginsberg, J.R., Griffiths, M.,
573 Holden, J., Kawanishi, K., Kinnaird, M., Laidlaw, R., 2001. The use of photographic
574 rates to estimate densities of tigers and other cryptic mammals. *Anim. Conserv.* 4(1):
575 75-79.
- 576 Carss, D., 1995. Foraging behaviour and feeding ecology of the otter *Lutra lutra*: a selective
577 review. *Hystrix It. J. Mamm.* 7: 179-194.
- 578 Chandler, R.B., Royle, J.A., 2013. Spatially Explicit Models for Inference About Density in
579 Unmarked or Partially Marked Populations. *Ann. Appl. Stat.* 7(2): 936-954.
- 580 Chatterjee, N., Nigam, P., Habib, B., 2020. Population density and habitat use of two
581 sympatric small cats in a central Indian reserve. *PLoS One* 15(6): p.e0233569.
- 582 Clevenger, A. P., Ford, A. T., Sawaya M. A., 2009. Banff wildlife crossings project:
583 Integrating science and education in restoring population connectivity across
584 transportation corridors. Final report to Parks Canada Agency, Radium Hot Springs,
585 British Columbia, Canada. Available from [http://arc-solutions.org/wp-](http://arc-solutions.org/wp-content/uploads/2012/03/Clevenger-et-al-2009-Banff-wildlife-crossings-project.pdf)
586 [content/uploads/2012/03/Clevenger-et-al-2009-Banff-wildlife-crossings-project.pdf](http://arc-solutions.org/wp-content/uploads/2012/03/Clevenger-et-al-2009-Banff-wildlife-crossings-project.pdf) [3
587 February 2020].
- 588 Corlatti, L., Sivieri, S., Sudolska, B., Giacomelli, S., Pedrotti, L., 2020. A field test of
589 unconventional camera trap distance sampling to estimate abundance of marmot
590 populations. *Wildlife Biol.* 2020(4): 1-11. doi: 10.2981/wlb.00652
- 591 Cusack, J.J., Swanson, A., Coulson, T., Packer, C., Carbone, C., Dickman, A.J., Kosmala, M.,
592 Lintott, C., Rowcliffe, J.M., 2015. Applying a random encounter model to estimate lion
593 density from camera traps in Serengeti National Park, Tanzania. *J. Wildl. Manag.* 79(6):
594 1014-1021.

- 596 Cutter, P., 2015. Fishing cat ecology: food habits, home ranges, habitat use and mortality in a
597 human-dominated landscape around Khao Sam Roi Yot, Peninsular Thailand. M. Sc.
598 thesis. Available from the University of Minnesota Digital Conservancy.
599 <https://hdl.handle.net/11299/174755>
- 600 de Silva, P.K., 1996. Food and feeding habits of the Eurasian Otter *Lutra lutra* L.(Carnivora,
601 Mustelidae) in Sri Lanka. J. South Asian Nat. Hist. 2(1): 81-90.
- 602 Dettori, E.E., Balestrieri, A., Zapata-Perez, V.M., Bruno, D., Rubio-Saura, N., Robledano-
603 Aymerich, F., 2021. Distribution and diet of recovering Eurasian otter (*Lutra lutra*)
604 along the natural-to-urban habitat gradient (river Segura, SE Spain). Urban Ecosyst.
605 2021: 1-10. <https://doi.org/10.1007/s11252-021-01109-3>
- 606 Delignette-Muller, M.L., Dutang, C., 2015. “fitdistrplus: An R Package for Fitting
607 Distributions.” J. Stat. Softw. 64(4): 1–34. <https://www.jstatsoft.org/v64/i04/>
- 608 Dissanayake, S.R.B., 1995. Avifaunal ecology with respect to different habitat types in the
609 Maduruoya National Park. M. Phil. thesis, Division of Zoology, Faculty of Natural
610 Sciences, The Open University of Sri Lanka, Nawala, Sri Lanka.
- 611 Dittus, W.P., 2017. The biogeography and ecology of Sri Lankan mammals point to
612 conservation priorities. Ceylon J. Sci. 46(special Issue): 33-64.
- 613 Engeman, R.M., Massei, G., Sage, M., Gentle, M.N., 2013. Monitoring wild pig populations:
614 a review of methods. Environ. Sci. Pollut. Res. 20(11): 8077-8091.
- 615 Fewster, R., S. Buckland, K. Burnham, D. Borchers, P. Jupp, Laake, J., Thomas, L., 2009.
616 Estimating the encounter rate variance in distance sampling. Biometrics 65: 225–236.

618 Ganguly, D., Adhya, T., 2020. How Fishing Cats *Prionailurus viverrinus* fish: Describing a
619 felid's strategy to hunt aquatic prey. bioRxiv. Available online at:
620 <https://doi.org/10.1101/2020.04.24.058925> Accessed 4 March 2021.

621 Gilbert, N. A., Clare, J. D. J., Stenglein, J. L., Zuckerberg, B., 2020. Abundance estimation of
622 unmarked animals based on camera-trap data. *Conserv. Biol.* 35(1): 88-100.

623 Glover-Kapfer, P., Soto-Navarro, C.A., Wearn, O.R., 2019. Camera-trapping version 3.0:
624 current constraints and future priorities for development. *Remote Sens. Ecol. Con.* 5(3):
625 209-223.

626 Gray, T.N.E., 2018. Monitoring tropical forest ungulates using camera-trap
627 data. *J. Zool.*, 305(3): 173-179.

628 Green, S.E., Rees, J.P., Stephens, P.A., Hill, R.A., Giordano, A.J., 2020. Innovations in
629 camera trapping technology and approaches: The integration of citizen science and
630 artificial intelligence. *Animals* 10(1): 132.

631 Hájková, P., Zemanová, B., Roche, K., Hájek, B., 2009. An evaluation of field and
632 noninvasive genetic methods for estimating Eurasian otter population size. *Conserv.*
633 *Genet.* 10(6): 1667-1681.

634 Haque, N.M., Vijayan, V., 1993. Food habits of the fishing cat *Felis viverrina* in Keoladeo
635 National Park, Bharatpur, Rajasthan. *J. Bombay Nat. Hist. Soc.* 90(1993): 498-500.

636 Hardouin, M., Searle, C.E., Strampelli, P., Smit, J., Dickman, A., Lobora, A.L., Rowcliffe,
637 J.M., 2021. Density responses of lesser-studied carnivores to habitat and management
638 strategies in southern Tanzania's Ruaha-Rungwa landscape. *PloS One* 16(3):
639 p.e0242293.

641 Harris, G.M., Butler, M.J., Stewart, D.R., Rominger, E.M., Ruhl, C.Q., 2020. Accurate
642 population estimation of Caprinae using camera traps and distance sampling. *Scientific*
643 *Sci. Rep.* 10(1): 17729. <https://doi.org/10.1038/s41598-020-73893-5>

644 Hoffmann, A., Decher, J., Rovero, F., Schaer, J., Voigt, C., Wibbelt, G., 2010. Field methods
645 and techniques for monitoring mammals. In: *Manual on field recording techniques and*
646 *protocols for all taxa biodiversity inventories and monitoring.* Eymann, J., Degreef, J.,
647 Hauser, C., Monje, J.C., Samyn, Y., Van den Spiegel D (Eds) Pensoft Publishers, Sofia,
648 Bulgaria. 482-529.

649 Hofmeester, T.R., Rowcliffe, J.M., Jansen, P.A., 2017. A simple method for estimating the
650 effective detection distance of camera traps. *Remote Sens. Ecol. Con.* 3(2): 81-89.

651 Howe, E. J., Buckland, S. T., Després-Einspenner, M.-L., Kühl, H.S., 2017. Distance
652 sampling with camera traps. *Methods Ecol. Evol.* 8(11): 1558–
653 1565. <https://doi.org/10.1111/2041-210X.12790>

654 Howe, E. J., Buckland, S. T., Després-Einspenner, M., Kühl, H. S., 2019. Model selection
655 with overdispersed distance sampling data. *Methods Ecol. Evol.* 10(1): 38-
656 47. <https://doi.org/10.1111/2041-210X.13082>

657 Hunter, L., 2019. *Carnivores of the world.* 2nd ed. Princeton University Press, Princeton,
658 New Jersey.

659 IUCN (International Union for Conservation of Nature). 1990. *IUCN Directory of South*
660 *Asian Protected Areas.* Gland, Switzerland and Cambridge, IUCN, UK. xxiv + 294.

661 IUCN (International Union for Conservation of Nature). 2021. *The IUCN red list of*
662 *threatened species.* Version 2021-1. Retrieved from <https://www.iucnredlist.org>

664 Jayaratne, C., Seneviratne S., 2020. Sri Lankan Jackal: The Island's True Wild Canid. *Loris*
665 29(2): 19-23.

666 Jayasekara, D., Kumara, P.K.P.M.P., Mahaulpatha, W.A.D., 2021. Mapping the vegetation
667 cover and habitat categorization of Maduru Oya and Horton Plains National Parks
668 using Landsat 8 (OLI) imagery to assist the ecological studies. *Wildlanka* 9(1): 122-135.

669 Jayasekara, E.G.D.P., Mahaulpatha, W.A.D., 2019. Abundance and Distribution of Family
670 Herpestidae in Three Protected Areas Representing Three Bioclimatic Zones in Sri
671 Lanka. Proceedings of the 6th International Conference on Multidisciplinary
672 Approaches (iCMA) 2019. Available at SSRN: <https://ssrn.com/abstract=3497273>

673 Jiménez, J., Nuñez-Arjona, J.C., Rueda, C., González, L.M., García-Domínguez, F., Muñoz-
674 Igualada, J., López-Bao, J.V., 2017. Estimating carnivore community
675 structures. *Sci. Rep.* 7(1): 1-10.

676 Johansson, Ö., Samelius, G., Wikberg, E., Chapron, G., Mishra, C., Low, M., 2020.
677 Identification errors in camera-trap studies result in systematic population
678 overestimation. *Sci. Rep.* 10(1): 1-10.

679 Kalle, R., 2013. Ecology of sympatric small carnivores in Mudumalai Tiger Reserve, Tamil
680 Nadu. Ph D. thesis, Saurashtra University, Rajkot, India. Available online
681 at : <http://hdl.handle.net/10603/15963> (Accessed: 18 June 2018).

682 Kalle, R., Ramesh, T., Sankar, K., Qureshi, Q., 2013. Observations of sympatric small
683 carnivores in Mudumalai Tiger Reserve, Western Ghats, India. *Small Carniv.*
684 *Conserv.* 49: 53-59.

685 Karanth, K.U., Nichols, J.D., Kumar, N.S., Hines, J.E., 2006. Assessing tiger population
686 dynamics using photographic capture–recapture sampling. *Ecology*, 87(11): 2925-2937.

688 Kitchener, A.C., Van Valkenburgh, B., Yamaguchi, N., 2010. Felid form and function. In:
689 Biology and Conservation of Wild Felids. MacDonald D.W., Loveridge, A.J (Eds)
690 Oxford University Press, Oxford. 83-106.

691 Kittle, A.M., Watson, A.C., 2018. Density of leopards (*Panthera pardus kotiya*) in Horton
692 Plains National Park in the Central Highlands of Sri Lanka. Mammalia 82(2):183-187.

693 Kittle, A.M., Watson, A.C., Fernando, T.S.P., 2017. The ecology and behaviour of a
694 protected area Sri Lankan leopard (*Panthera pardus kotiya*)
695 population. Trop. Ecol. 58(1): 71-86.

696 Lanszki, J., Hidas, A., Szentes, K., Révay, T., Lehoczky, I., Weiss, S., 2008. Relative spraint
697 density and genetic structure of otter (*Lutra lutra*) along the Drava River in
698 Hungary. Mamm. Biol. 73(1): 40-47.

699 Luo, G., Wei, W., Dai, Q., Ran, J., 2020. Density Estimation of Unmarked Populations Using
700 Camera Traps in Heterogeneous Space. Wildl. Soc. Bull. 44(1): 173-181.

701 Manzo, E., Bartolommei, P., Rowcliffe, J.M., Cozzolino, R., 2012. Estimation of population
702 density of European pine marten in central Italy using camera
703 trapping. Acta Theriol. 57(2): 165-172.

704 Marques, T.A., Thomas, L., Martin, S.W., Mellinger, D.K., Ward, J.A., Moretti, D.J., Harris,
705 D., Tyack, P.L., 2013. Estimating animal population density using passive
706 acoustics. Biol. Rev. 88(2): 287-309.

707 Meek, P.D., Ballard, G., Falzon, G., Williamson, J., Milne, H., Farrell, R., Stover, J., Mather-
708 Zardain, A.T., Bishop, J.C., Cheung, E.K.W., Lawson, C.K., 2020. Camera trapping
709 technology and related advances: Into the new millennium. Aust. J. Zool. 40(3): 392-
710 403.

712 Meek, P.D., Fleming, P., Ballard, G., Banks, P., Claridge, A.W., Sanderson, J., Swann, D.
713 (Eds) 2014. Camera trapping: wildlife management and research (Vol. 2). Csiro
714 Publishing, Melbourne, Australia.

715 Meek, P.D., Zewe, F., Falzon, G., 2012. Temporal activity patterns of the swamp rat (*Rattus*
716 *lutreolus*) and other rodents in north-eastern New South Wales, Australia. *Aust.*
717 *Mammal.* 34(2): 223-233.

718 Mishra, R., Basnet, K., Amin, R., Lamichhane, B.R., 2018. Fishing Cat *Prionailurus*
719 *viverrinus* Bennett, 1833 (Carnivora: Felidae) distribution and habitat characteristics in
720 Chitwan National Park, Nepal. *J. Threat. Taxa.* 10(11): 12451-12458.

721 Miththapala, S., 2018. An overview of the wild cats of Sri Lanka Part II. *Loris* 28(3): 7-47.

722 MOE (Ministry of Environment), 2012. The National Red List 2012 of Sri Lanka;
723 Conservation Status of the Fauna and Flora. Ministry of Environment, Colombo, Sri
724 Lanka. viii + 476.

725 Moeller, A.K., Lukacs, P.M., Horne, J.S., 2018. Three novel methods to estimate abundance
726 of unmarked animals using remote cameras. *Ecosphere* 9(8): e02331.

727 MoMD&E (Biodiversity Secretariat, Ministry of Mahaweli Development and Environment),
728 2019. Biodiversity Profile - Sri Lanka, Sixth National Report to the Convention on
729 Biological Diversity, Biodiversity Secretariat, Ministry of Mahaweli Development and
730 Environment, Sri Lanka.

731 Morrison, M.L., 2013. Wildlife restoration: techniques for habitat analysis and animal
732 monitoring (Vol. 1). Island Press, Washington, DC.

734 Mukherjee, S., Appel, A., Duckworth, J.W., Sanderson, J., Dahal, S., Willcox, D.H.A.,
735 Herranz Muñoz, V., Malla, G., Ratnayaka, A., Kantimahanti, M., Thudugala, A., 2016.
736 *Prionailurus viverrinus*. The IUCN Red List of Threatened Species 2016:
737 eT18150A50662615.

738 Nakashima, Y., Fukasawa, K., Samejima, H., 2018. Estimating animal density without
739 individual recognition using information derivable exclusively from camera
740 traps. *J Appl Ecol.* 55(2): 735-744.

741 Nekaris, K. A. I. 2003. Distribution and behaviour of three small wild cats in Sri Lanka. *Cat*
742 *News* 38(1): 30-32.

743 O'Brien, T.G., 2011. Abundance, density and relative abundance: a conceptual framework. In:
744 *Camera traps in animal ecology*. Springer, Tokyo. 71-96.

745 Pal, R., Bhattacharya, T., Qureshi, Q., Buckland, S. T., Sathyakumar, S., 2021. Using
746 distance sampling with camera traps to estimate the density of group-living and solitary
747 mountain ungulates. *Oryx*. Cambridge University Press: 1–9. doi:
748 10.1017/S003060532000071X.

749 Palei, H.S., Palei, N.C., Rath, B.P., Mishra, A.K., 2019. Records of the globally threatened
750 Rusty-spotted Cat in Odisha, India. *Nat. Conserv. Res.* 4(3): 112-116.

751 Palencia, P., 2020. trappingmotion: Integrate camera-trapping in movement and behavioural
752 studies. R package version 0.1.1. <https://github.com/PabloPalencia/trappingmotion>

753 Palencia, P., Fernández-López, J., Vicente, J., Acevedo, P., 2021a. Innovations in movement
754 and behavioural ecology from camera traps: day range as model
755 parameter. *Methods Ecol. Evol.* 12(7): 1201-1212. [https://doi.org/10.1111/2041-](https://doi.org/10.1111/2041-210X.13609)
756 [210X.13609](https://doi.org/10.1111/2041-210X.13609)

758 Palencia, P., Rowcliffe, J.M., Vicente, J., Acevedo, P., 2021b. Assessing the camera trap
759 methodologies used to estimate density of unmarked populations. J. Appl.
760 Ecol. 2021(00): 1–10. <https://doi.org/10.1111/1365-2664.13913>

761 Palencia, P., Vicente, J., Barroso, P., Barasona, J.Á., Soriguer, R.C., Acevedo, P., 2019.
762 Estimating day range from camera-trap data: the animals' behaviour as a key
763 parameter. J. Zool. 309(3): 182-190.

764 Parker, I., Lopez, R., Silvy, N., Davis, D., Cathey, J., 2012. Alternative methodology for
765 handling and marking meso-mammals for short-term research. Wildl. Biol. Pract. 8(2):
766 20-25.

767 Pfeffer, S.E., Spitzer, R., Allen, A.M., Hofmeester, T.R., Ericsson, G., Widemo, F., Singh,
768 N.J., Cromsigt, J.P., 2018. Pictures or pellets? Comparing camera trapping and dung
769 counts as methods for estimating population densities of ungulates. Remote Sens. Ecol.
770 Con. 4(2): 173-183.

771 Punyawardena B.V.R., 2020. Climate. In: Mapa R., (Ed). The Soils of Sri Lanka. World Soils
772 Book Series, Springer, Cham. 13-22. https://doi.org/10.1007/978-3-030-44144-9_2

773 Quaglietta, L., Hájková, P., Mira, A., Boitani, L., 2015. Eurasian otter (*Lutra lutra*) density
774 estimate based on radio tracking and other data sources. Mammal Res. 60(2): 127-137.

775 R Core Team, 2013. R: A language and environment for statistical computing. R Foundation
776 for Statistical Computing, Vienna, Austria. <http://www.r-project.org>

777 Rademaker, M., Meijaard, E., Semiadi, G., Blokland, S., Neilson, E.W., Rode-Margono, E.J.,
778 2016. First ecological study of the Bawean warty pig (*Sus blouchi*), one of the rarest
779 pigs on earth. PLoS One, 11(4):.e0151732.

- 781 Ramsey, D.S., Caley, P.A., Robley, A., 2015. Estimating population density from presence–
782 absence data using a spatially explicit model. *J. Wildl. Manag.* 79(3): 491-499.
- 783 Rich, L.N., Miller, D.A., Muñoz, D.J., Robinson, H.S., McNutt, J.W., Kelly, M.J., 2019.
784 Sampling design and analytical advances allow for simultaneous density estimation of
785 seven sympatric carnivore species from camera trap data. *Biol. Conserv.* 233(2019):
786 12-20.
- 787 Roemer, G.W., Gompper, M.E., Van Valkenburgh, B., 2009. The ecological role of the
788 mammalian mesocarnivore. *BioScience.* 59(2):165-173.
- 789 Romairone, J., Jiménez, J., Luque-Larena, J.J., Mougeot, F., 2018. Spatial capture-recapture
790 design and modelling for the study of small mammals. *PLoS One*, 13(6): e0198766.
- 791 Romero, R., Guitián, J., 2017. Food and feeding habits of Eurasian otter, *Lutra lutra*, and
792 American mink, *Neovison vison*, in an Atlantic island of northwest Spain. *J. Vertebr.*
793 *Biol.* 66(2): 117-125.
- 794 Rovero F., De Luca, D.W., 2007. Checklist of mammals of the Udzungwa Mountains of
795 Tanzania. *Mammalia* 71: 47–55.
- 796 Rovero, F., Marshall, A.R., 2009. Camera trapping photographic rate as an index of density
797 in forest ungulates. *J. Appl. Ecol.* 46(5): 1011-1017.
- 798 Rovero, F., Zimmermann, F., Berzi, D., Meek, P., 2013. "Which camera trap type and how
799 many do I need?" A review of camera features and study designs for a range of wildlife
800 research applications. *Hystrix It. J. Mamm.* 24(2): 148–156.
- 801 Rowcliffe, J. M., 2019. *activity: Animal activity statistics*. R package version
802 1.3. <https://CRAN.R-project.org/package=activity>

804 Rowcliffe, J.M., Carbone, C., Jansen, P.A., Kays, R., Kranstauber, B., 2011. Quantifying the
805 sensitivity of camera traps: an adapted distance sampling
806 approach. *Methods Ecol. Evol.* 2(5): 464-476.

807 Rowcliffe, J.M., Carbone, C., Kays, R., Kranstauber, B., Jansen, P.A., 2012. Bias in
808 estimating animal travel distance: the effect of sampling
809 frequency. *Methods Ecol. Evol.* 3(4): 653-662.

810 Rowcliffe, J.M., Field, J., Turvey, S.T., Carbone, C., 2008. Estimating animal density using
811 camera traps without the need for individual recognition. *J. Appl. Ecol.* 45(4): 1228-
812 1236.

813 Rowcliffe, J.M., Jansen, P.A., Kays, R., Kranstauber, B., Carbone, C., 2016. Wildlife speed
814 cameras: measuring animal travel speed and day range using camera traps. *Remote*
815 *Remote Sens. Ecol. Con.* 2(2): 84-94.

816 Rowcliffe, J.M., Kays, R., Kranstauber, B., Carbone, C., Jansen, P.A., 2014. Quantifying
817 levels of animal activity using camera trap data. *Methods Ecol. Evol.* 5(11): 1170-1179.

818 Royle, J.A., Nichols, J.D., 2003. Estimating abundance from repeated presence-absence data
819 or point counts. *Ecology* (84): 777-790.

820 Royle, J.A., 2004. N-mixture models for estimating population size from spatially replicated
821 counts. *Biometrics* 60(1): 108-115.

822 Royle, J.A., Chandler, R.B., Gazenski, K.D., Graves, T.A., 2013. Spatial capture–recapture
823 models for jointly estimating population density and landscape
824 connectivity. *Ecology* 94(2): 287-294.

826 Ruiz-Olmo, J., Saavedra, D., Jiménez, J., 2001. Testing the surveys and visual and track
827 censuses of Eurasian otters (*Lutra lutra*). J. Zool. 253(3): 359-369.

828 Šálek, M., Červinka, J., Banea, O.C., Krofel, M., Čirović, D., Selanec, I., Penezić, A., Grill,
829 S., Riegert, J., 2014. Population densities and habitat use of the golden jackal (*Canis*
830 *aureus*) in farmlands across the Balkan Peninsula. Eur. J. Wildl. Res. 60(2):193-200.

831 Šálek, M., Kreisinger, J., Sedláček, F., Albrecht, T., 2010. Do prey densities determine
832 preferences of mammalian predators for habitat edges in an agricultural
833 landscape?. Landsc. Urban Plan. 98(2): 86-91.

834 Sanderson, J.G., Trolle, M., 2005. Monitoring elusive mammals: unattended cameras reveal
835 secrets of some of the world's wildest places. Am. Sci. 93(2): 148-155.

836 Santiapillai, C., De Silva, M., Dissanayake, S.R.B., 2000. The status of mongooses (Family:
837 Herpestidae) in Ruhuna National Park, Sri Lanka. J. Bombay Nat. Hist. Soc. 97(2):
838 208-214.

839 Sathiyaselvam, P., Satyanarayana, E., Kathula, T., 2016. Current status and threats to fishing
840 cat *Prionailurus viverrinus* (Bennett, 1833) in Godavari Mangroves, Andhra Pradesh,
841 India. Indian For. 142(10): 950-832.

842 Scar.lk., 2021. SCAR. [online] Available at: <<https://scar.lk/>> [Accessed 12 March 2021].

843 Sheftel, B.I., 2018. Methods for estimating the abundance of small mammals. Russ J
844 Ecol. 3(3): 1-21.

845 Silveira, L., Jacomo, A.T., Diniz-Filho, J.A.F., 2003. Camera trap, line transect census and
846 track surveys: a comparative evaluation. Biol. Conserv. 114(3): 351-355.

848 Srbek-Araujo, A.C., Chiarello, A.G., 2005. Is camera-trapping an efficient method for
849 surveying mammals in Neotropical forests? A case study in south-eastern
850 Brazil. *J. Trop. Ecol.* 21(1): 121-125.

851 TEAM Network., 2011. Terrestrial Vertebrate Protocol Implementation Manual, v. 3.1.
852 Tropical Ecology, Assessment and Monitoring Network, Center for Applied
853 Biodiversity Science, Conservation International, Arlington, VA, USA.

854 Thomas, L., Buckland, S.T., Rexstad, E.A., Laake, J.L., Strindberg, S., Hedley, S.L., Bishop,
855 J.R., Marques, T.A., Burnham, K.P., 2010. Distance software: design and analysis of
856 distance sampling surveys for estimating population size. *J. Appl. Ecol.* 47(1): 5-14.

857 Tobler, M.W., Carrillo-Percegué, S.E., Powell, G., 2009. Habitat use, activity patterns and
858 use of mineral licks by five species of ungulate in south-eastern Peru. *J. Trop. Ecol.*
859 25(3): 261-270.

860 Wald, A., Wolfowitz, J., 1940. On a test whether two samples are from the same
861 population. *Ann. Math. Stat.* 11(2): 147– 162. <http://www.jstor.org/stable/2235872>

862 Webb, E.L., Choo, Y.R., Kudavidanage, E.P., Amarasinghe, T.R., Bandara, U.G.S.I.,
863 Wanninayaka, W.A.C.L., Ravindrakumar, P., Nimalrathna, T.S., Liang, S.H., Chua,
864 M.A.H., 2020. Leopard activity patterns in a small montane protected area highlight the
865 need for integrated, collaborative landscape conservation. *Glob. Ecol. Conserv.* 23:
866 e01182.

867 Weerakoon, D.K., Goonatilake, W.D.A., 2006. Taxonomic status of the mammals of Sri
868 Lanka. *Fauna of Sri Lanka: Status of Taxonomy, Research and Conservation*, IUCN Sri
869 Lanka, Colombo. 216-231.

- 871 Weerakoon, D.K., 2012. The taxonomy and conservation status of mammals in Sri Lanka. In:
872 The National Red List 2012 of Sri Lanka; Conservation Status of the Fauna and Flora.
873 Weerakoon, D.K., Wijesundara S. (Eds). Ministry of Environment, Colombo, Sri Lanka.
874 134-144.
- 875 Wijesinghe, M.R., 2006. Ecological traits of endemic small mammals in rainforests of Sri
876 Lanka, and their implications for conservation. In: Bambaradeniya, C.N.B. (Ed). The
877 Fauna of Sri Lanka: Status of Taxonomy, Research, and Conservation, IUCN, Colombo,
878 Sri Lanka. 232-234.
- 879 Wijeyeratne, G.D., 2008. A photographic Guide to Mammals of Sri Lanka. New UK:
880 Holland Publishers.
- 881 Zimmermann F., Molinari-Jobin A., Ryser A., Breitenmoser-Würsten Ch., Pesenti E.,
882 Breitenmoser U., 2011. Status and distribution of the lynx (*Lynx lynx*) in the Swiss Alps
883 2005–2009. Acta Biol. Slov. 54: 74–84.

Table 1: Additional parameters derived for density calculations. ADD: average detection distance; EDD: effective detection distance; Speed of animal movement; Activity pattern (sD = strictly diurnal; sN = strictly nocturnal; mD = mainly diurnal; mN = mainly nocturnal; Activity level: the proportion of the day a species is active); Day range: daily distance travelled; IUCN status (IUCN, 2021).

Species	ADD (m)	EDD (m)	Movement Speed (km/h)	Activity Pattern	Activity level	Day range (km/day)	IUCN status (Global)
Fishing cat <i>Prionailurus viverrinus</i>	2.54	2.75	0.72	mN	0.461	7.96	VU
Rusty-spotted cat <i>Prionailurus rubiginosus</i>	1.9	-	-	mN	-	-	NT
Jungle Cat <i>Felis chaus</i>	2.62	-	-	mN	-	-	LC
Ring-tailed civet <i>Viverricula indica</i>	2.84	3.19	1.02	sN	0.288	7.05	LC
Golden palm civet <i>Paradoxurus zeylonensis</i>	3.01	2.89	0.86	sN	0.161	3.34	LC
Stripe-necked mongoose <i>Urva vitticollis</i>	3.11	3.35	1.22	sD	0.288	8.39	LC
Ruddy mongoose <i>Urva smithii</i>	2.91	3.47	1.84	sD	0.390	17.22	LC
Brown mongoose <i>Urva fuscus</i>	2.92	-	-	sD	-	-	LC
Otter <i>Lutra lutra</i>	4.07	4.92	3.42	mN	0.353	28.97	NT
Golden jackal <i>Canis aureus</i>	3.35	3.97	3.10	mD	0.419	31.13	LC

Table 2: Density estimates of meso-mammal carnivore species in MONP using three analytical methods: REM, conventional CTDS and bootstrap CTDS(b) density estimates. (Density is given as individuals/km², LCL=Lower Confidence Limit; UCL=Upper Confidence Limit; %CV=percent Coefficient of Variation). CTDS model function: half normal - (hn), hazard rate - (hr). RAI: Relative Abundance Index.

Species	RAI	Density Estimate Method	Density (individuals per km ²)			
			Estimate	LCL	UCL	% CV
Fishing cat <i>P. viverrinus</i>	3.11	REM	1.54	0.82	2.39	29.0
		CTDS(b) (hr)	1.13	0.45	2.90	50.1
		CTDS (hr)	0.90	0.51	1.60	29.8
Rusty-spotted cat <i>P. rubiginosus</i>	0.12	-	-	-	-	-
Jungle cat <i>F. chaus</i>	0.15	-	-	-	-	-
Ring-tailed civet <i>V. indica</i>	3.47	REM	2.28	1.13	3.57	35.8
		CTDS(b) (hr)	1.91	1.03	3.55	31.9
		CTDS (hr)	1.69	1.09	2.63	22.6
Golden palm civet <i>P. zeylonensis</i>	1.32	REM	1.69	1.17	2.29	20.8
		CTDS(b) (hn)	0.80	0.32	1.98	48.6
		CTDS (hn)	0.97	0.42	2.24	44.2
Stripe-necked mongoose <i>U. vitticollis</i>	1.53	REM	0.75	0.47	1.06	23.0
		CTDS(b) (hr)	0.62	0.32	1.22	34.9
		CTDS (hr)	0.56	0.34	0.93	26.1
Ruddy mongoose <i>U. smithii</i>	8.88	REM	2.19	1.48	2.95	21.3
		CTDS(b) (hr)	2.32	1.37	3.93	27.1
		CTDS (hr)	2.23	1.40	3.56	23.9
Brown mongoose <i>U. fuscus</i>	0.29	-	-	-	-	-
Otter <i>L. lutra</i>	1.35	REM	0.15	0.05	0.28	45.9
		CTDS(b) (hr)	0.16	0.07	0.36	45.0

934		CTDS (hr)	0.15	0.05	0.47	61.1	
935	Golden jackal	REM	0.17	0.07	0.27	39.1	
936	<i>C. aureus</i>	1.32	CTDS(b) (hn)	0.16	0.60	0.42	51.67
937			CTDS (hn)	0.16	0.07	0.40	48.5
938							

940 **Figure 1:** Map of Maduru Oya National Park with the study area and camera station
941 locations. Location of the park in the map of Sri Lanka is also shown

942 **Figure 2:** The distance grid superimposed on camera trap capture frame to estimate distances

943 **Figure 3:** (A) Rusty-spotted cat, (B) Stripe-necked mongoose (C) Fishing cat (D) Ring-tailed
944 civet and (E) Golden palm civet captured in our camera traps

945 **Figure 4:** Activity patterns of fishing cat, ring-tailed civet, ruddy mongoose and golden
946 jackal in MONP, as captured by distributions of camera-trap records. Black steps are
947 observed frequencies, and curves are fitted circular kernel distributions

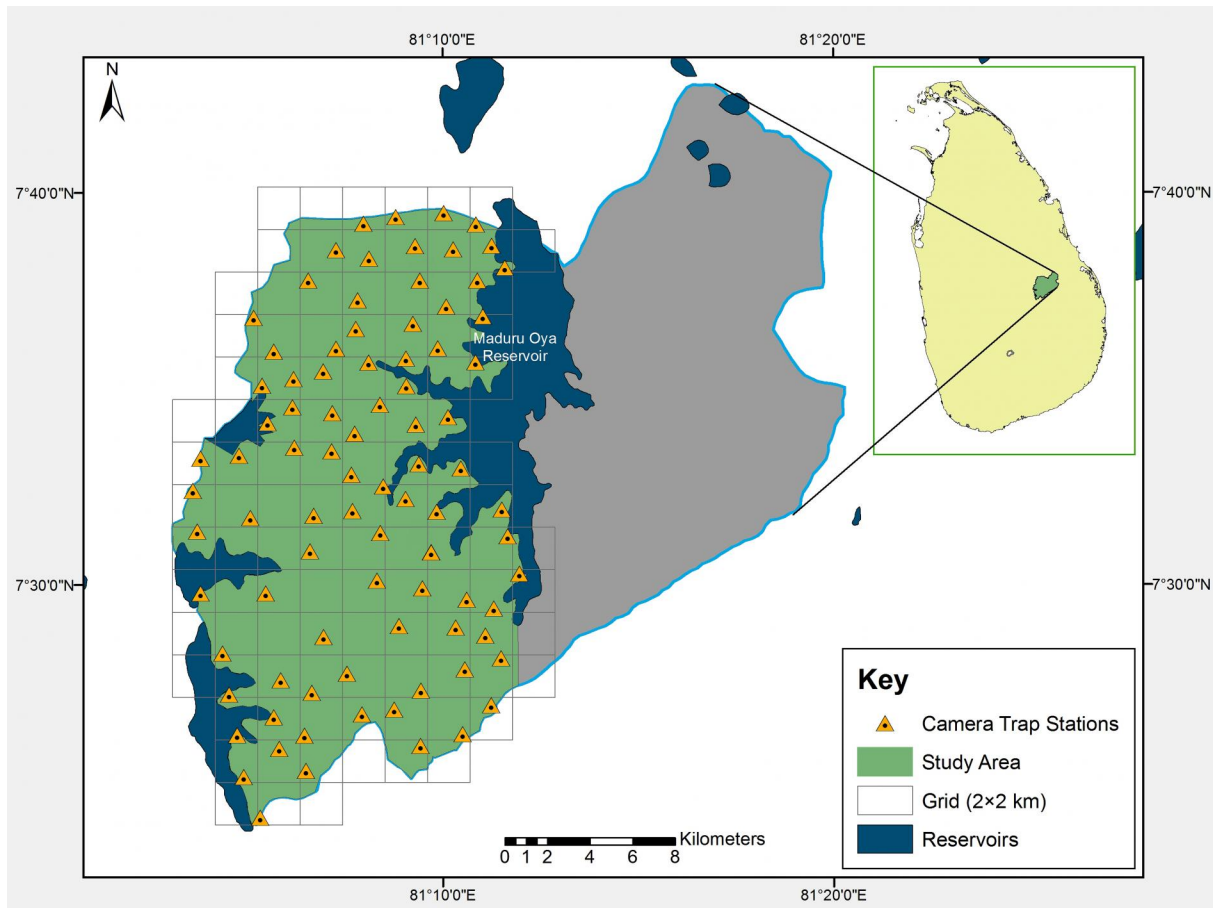


Figure 1: Map of Maduru Oya National Park with the study area and camera station locations. Location of the park in the map of Sri Lanka is also shown.

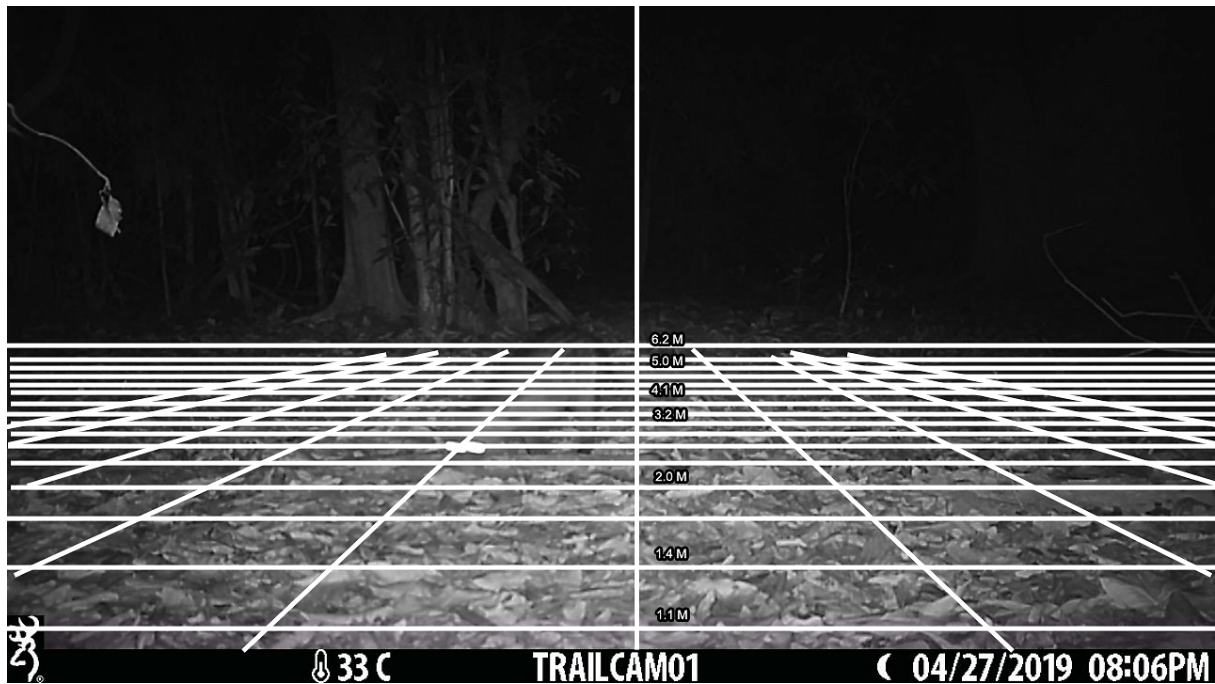


Figure 2: The distance grid superimposed on camera trap capture frame to estimate distances

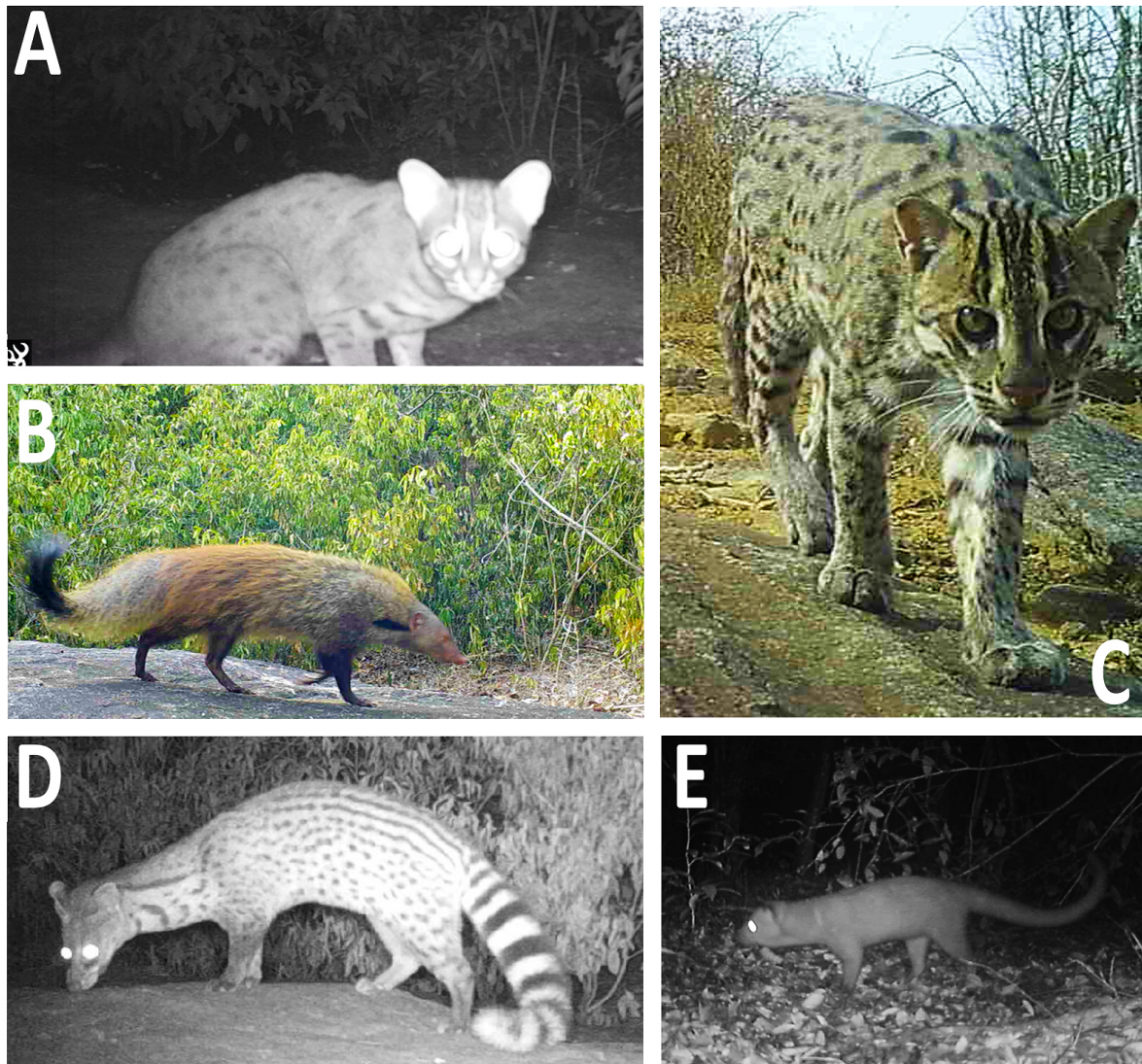


Figure 3: (A) Rusty-spotted cat, (B) Stripe-necked mongoose a (C) Fishing cat (D) Ring-tailed civet, and (E) Golden palm civet captured in our camera traps

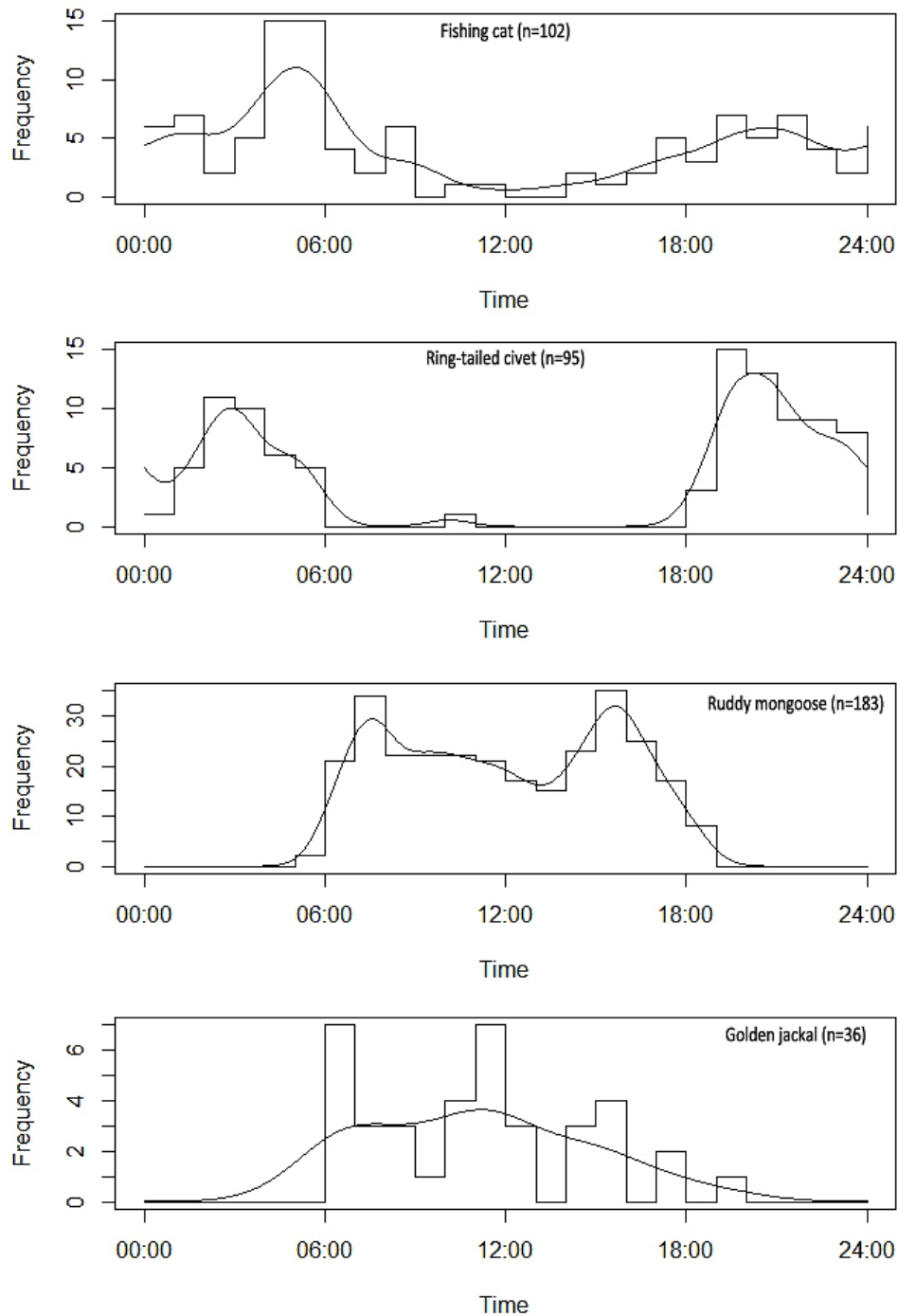


Figure 4: Activity patterns of fishing cat, ring-tailed civet, ruddy mongoose and golden jackal in MONP, as captured by distributions of camera-trap records. Grey steps are observed frequencies, and black curves are fitted circular kernel distributions

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Figure 1: Map of Maduru Oya National Park with the study area and camera station locations. Location of the park in the map of Sri Lanka is also shown.

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Figure 4: Activity patterns of fishing cat, ring-tailed civet, ruddy mongoose and golden jackal in MONP, as captured by distributions of camera-trap records. Grey steps are observed frequencies, and black curves are fitted circular kernel distributions

Supplementary Online Material

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