The promise of molecular identification of fungi to overcome the global challenges in plant biosecurity

D. Udayanga

Department of Bio-systems Technology, Faculty of Technology, University of Sri Jayewardenepura dudayanga@sjp.ac.lk

Abstract

Phytopathogenic fungi are a highly diverse group of eukaryotes with significant impact on natural ecosystems and cultivated lands, causing diseases on crops, ornamentals and forest trees. Historical outbreaks of invasive fungal pathogens on staple crops have caused severe food security threats to human populations. Therefore, accurate identification of pathogenic fungi has a great impact on global plant biosecurity and is considered to be crucial in this era of global exchange of horticultural products and plant germplasm. Although morphological features have traditionally been used in fungal identification, the past two decades have witnessed revolutionary changes with the implementation of DNA sequence data. The nuclear ribosomal internal transcribed spacer region (ITS), the standard fungal DNA barcode, is extensively used for a quick and often approximate identification. However, due to limitations of single DNA barcodes in fungal species discrimination and establishment of evolutionary relationships. multiple genomic loci have been utilized. The identification of fungi in plant biosecurity and quarantine practices are often based on morphology and tentative taxonomic assignments have increased the risk to regional and global plant biosecurity. This mini-review summarizes (1) the significance of precise naming of pathogenic fungi; (2) the incorporation of molecular data in the identification of emerging phytopathogens with a few examples; and (3) the need for paradigm shifts in global plant biosecurity practices. In addition, we urge the relevant agencies of the countries lacking organized plant biosecurity practices to recognize the need to confront the potential threats on their staple crops, ornamentals and forest trees and support appropriate for DNA-aided identification research and classification, and application of accurate names to high priority phytopathogens and emerging species. This will enable effective disease surveillance, prevent unnecessary trade restrictions for plantbased products and enhance biosecurity and quarantine efforts.

Keywords: Biological invasions, DNA barcoding, Emerging pathogens, Molecular phylogeny, Plant protection

Introduction

The kingdom of fungi constitutes a large, diverse group of eukaryotic, heterotrophic, ubiquitous microorganisms that play a significant role in human, animal and plant health, food security and ecosystem resilience on earth (Hibbett et al. 2007; Tedersoo et al. 2014). They are the dominant causal agents of a wide range of plant diseases (Udayanga et al. 2011; Dean et al. 2012). Saprotrophic fungi feed on dead plant tissues and parasitic fungi primarily live on or within living plants, causing various disease symptoms, whereas endophytic and mycorrhizal fungi exhibit a mutualistic association without causing apparent symptoms (McDonald & Stukenbrock 2016; Doehlemann et al. 2017).

Fungal diseases are threatening many commercially and socially valuable crops including cereals, fruits, vegetables, pulses, plantation and cash crops as well as ornamentals and forest trees (Park et al. 2000; Rossman et al. 2007). Emerging plant diseases, which may either be previously known or have recently shifted geographic regions or associated hosts have become a global challenge in crop biosecurity. The emergence of plant pathogens can be partly due to climate change, as well as the rapid development of international trade of horticultural products and other living plants (Elad & Pertot 2014). These emerging diseases are not only important in global crop production, but also pose severe risks on a local level, especially in export agriculture, plantation industry and small farms in developing countries. Invasive microorganisms, the species whose introduction or spread threatens human. animal, plant health, agricultural, environmental security or the economy are also one of the growing threats for plant biosecurity, human and animal health, and export agriculture worldwide (Paini et al. 2016; Stricker et al. 2016). The current inefficiencies of fungal identification, lack of expertise as well as the minimum implementation of molecular data have aggravated the issues pertinent to global food security (Crous et al. 2016; McDonald & Stukenbrock 2016).

Some of the widespread fungal diseases are major threats to the most important crops such as rice, wheat and maize upon which humanity depends, whereas some of them may cause significant losses of crops that are the main means of revenue for some countries or serious threats to biodiversity (Couch & Kohn 2002; Godfrey et al. 2016). For instance, the rice blast pathogen, Pyricularia oryzae, formerly known as Magnaporthe oryzae is the most destructive pathogen of rice, the widely consumed staple food for a large part of the world's human population (Couch & Kohn 2002). Severe outbreaks of the rice blast disease are a recurrent problem in all rice-growing regions of the world where the disease is found to be difficult to control (Kahn et al. 2016; Nalley, 2016). Thus, rice blast that destroys up to 30% of the world's rice crop in each year is considered as a significant economic and humanitarian issue, especially in Asia (Savary et al; 2000; Saleh et al. 2014). Similarly, the coffee rust disease caused by the fungus, Hemileia vastatrix, has plagued coffee growers globally for more than a century (McCook 2006; Vandermeer et al. 2010, 2018). The fungus which is found to be with an East African origin turned into a destructive pathogen in the major coffee growing regions of Sri Lanka (then called Ceylon in 1867 and had ruined completely the celebrated coffee production in the Central Highlands of the country. After more than a century, the same fungus hit the renowned coffeeproducing regions in Central America, causing a cluster of outbreaks since 2008. The succession of coffee rust outbreaks, termed "the big rust" by some biologists, begun in 2008 from Colombia, and then moved northward in 2012-2013, causing significant losses in parts of Central America and Mexico (McCook & Vandermeer 2015). Then it was reportedly shifted southward in 2014-2015 causing severe losses in Ecuador and Peru and again in 2018-2019 in Colombia, Honduras and throughout the region leading a massive hit on coffee-based economy (Ehrenbergerová et al. 2018).

Traditional species identification of fungi involved discovery, morphological descriptions, artificial system of classification and naming of fungal species (Rossman & Palm-Hernández 2008). Similarly, morphological identification has been implemented for fungi in almost all phytopathological studies as well as to detect pathogens in biosecurity applications and quarantine (Burgess et al. 2016). In morpho-species recognition, the macroscopic and microscopic fungal structures of the sexual and asexual morphs of were compared using available taxonomic literature and preserved voucher specimens in fungal herbaria, which has been the basis for traditional system of identification (Cai et al. 2011; Hyde et al. 2013). The same methodical approach has been implemented in biosecurity and quarantine situations, primarily by authorities and specialists working on fungal identifications around the world. Nevertheless, the fungal identification solely based on morphology leads to numerous difficulties and failures, especially the species level classification (Crous et al. 2015; Raja et al. 2017). In some highly diverse genera of fungi, the species delimitation has been impossible even for well-experienced mycologists, trained to work on fungal identification by profession. Morphological crypsis, plasticity and limitations of distinguishing morphological characters among species often result misidentifications (Udayanga et al. 2011). Moreover, the pleomorphic fungi, which comprise two entirely dissimilar asexual and sexual morphs in the life cycle of the same fungus has confused early mycologists and were even given different scientific names until recently. Therefore, the phenotypic characters alone in fungi has been deemed to be obsolete for many of the genera. These common drawbacks of morphological species identification can be due to evolutionary consequences, including hybridization, cryptic speciation, and convergent evolution. As a consequence of failures in fungal morpho-species recognition, different species recognition criteria, including physiological, biological and biochemical species recognition used before criteria were the molecular identification of fungi become prominent (Udayanga et al. 2011; Hibbett et al. 2016). In this review, we consider the necessity for accurate identification, applications of molecular data in species-level fungal identifications with few recent examples. We urge for international agreements for fundamental changes in biosecurity and quarantine practices pertinent to trade-associated fungal plant pathogens based on standardized DNA-aided identification systems.

Significance of Precise Naming of Pathogenic Fungi

Among all taxa of the kingdom fungi, plant pathogens represent a relatively less number of species, however much diverse group of organisms (Figure 1). Scientific names of pathogenic fungi are the key to all accumulated knowledge and those are the foundation for effective communication (Hawksworth 2001, 2015; Rossman et al. 2016). The names of fungi can be used to access the specific, scattered knowledge of the species identification, classification, geographic distribution, biochemistry and bioprospecting, biosecurity and quarantine, ecology, pathogenicity, virulence and fungal disease surveillance.

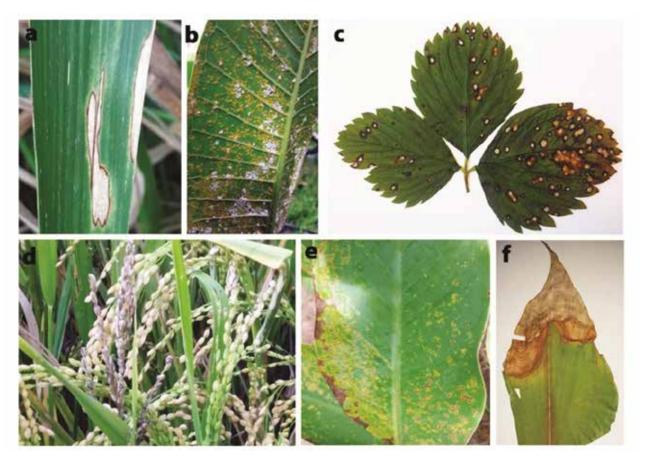


Figure 1: Diseases of a few economically important crops and ornamental plants.

a) Leaf spot of *Oryza sativa* (rice) –*Curvularia* sp. ; b) Rust on *Plumeria* (Frangipani) – *Coleosporium plumeriae* ; c) Purple leaf spot of *Fragaria* (strawberry)–*Ramularia grevilleana* ; d) Seed infections of rice –*Periconia* sp.; e) Rust on *Canna–Pucciniațhaliae* ; f) Leaf blight of *Cucurma* (Turmeric) –*Diaporthe* sp.

Specifically, in biosecurity and quarantine, the name of the fungus of interest is paramount in the process of interception procedures or quarantine actions. For instance, the coffee rust pathogen, *Hemileia vastatrix* has been associated with coffee plant which is economically and socially important crop, for over 150 years (Cressey 2013; Avelino et al. 2015). The information linked with this scientific name, *Hemileia vastatrix* for decades, scattered globally, including disease reports, biological literature, specimens in fungal herbaria and intercepted collections by biosecurity authorities. In the molecular era, numerous nucleotide sequence data as well as whole genome shotgun sequence data available in GenBank which provide crucial biological information associated with the fungus, and all facts are unified by this name (Carvalho et al. 2014).

The myrtle rust fungus, *Austropuccinia psidii* is considered to be an invasive species which has caused devastating effects in Australia in late 2018

and early 2019, since it has been first detected in Australia in 2010 on the NSW central coast (Morin et al. 2012). The myrtle rust formerly known as Puccinia psidii originates from South America and later become invasive infecting on several genera of Myrtaceae (e.g., Myrtle, pohutukawa, bay rum tree, clove, guava, acca) in Australia, causing emergency national action plans to mitigate the outbreak in January 2019. However, the taxonomic position and meaningful name for myrtle rust pathogen had been unclear until recently. Based on molecular phylogenetic analyses, Beenken (2017) showed that, it does not belong in the genus Puccinia, thus the new genus Austropuccinia was introduced to accommodate the myrtle rust fungus. Having a meaningful name for a fungus with greater impact on continent's major threat to biodiversity, allowed scientists to access more data and natural history information of the fungus that will be helpful to combat against this deleterious pathogen. This information will allow more precise risk assessments in support of scientifically sound plant quarantine decisions and policies that sustain world trade of agricultural commodities, while protecting the plant resources of individual countries.

According to the previous International Code of Botanical Nomenclature (ICBN), Article 59 permitted the application of different names to the different morphs of the same fungus, depending on whether a sexual (teleomorphic) or asexual (anamorphic) reproduction was involved (Norvell 2011). This dual nomenclature system, which allowed two or more names for the same species had resulted in significant failures in the consistency of identifying and naming species and no doubt on biosecurity and quarantine lists (Wingfield et al. 2012). Following the bold decisions taken by mycologists in "one-fungus one name" symposium and subsequent publication of the Amsterdam declaration of fungal nomenclature (Hawksworth et al. 2011), the International Botanical Congress held in Melbourne in 2011 (IBC-2011) approved several key resolutions that will affect fungal nomenclature (Norvell 2011; Taylor 2011). Accordingly, all

legitimate fungal names published prior to January 1, 2013 compete equally for priority, and the sole correct name is now the earliest legitimate name, regardless of the life history state of the type (Hawksworth 2011). However, this long awaited decision initially shocked plant pathological community who had their favorite names for different fungi, later embraced by them adjusting to more meaningful system in naming species (Geiser et al. 2013; Rossman et al. 2016). The changes of botanical code are widely implemented later in order to establish precise naming of fungi with greater importance in biosecurity in conjunction with the molecular phylogenetic results.

Incorporation of Molecular Data in the Identification of Emerging Phytopathogens with a Few Examples

The precise identification and molecular systematics fungi heavily utilize DNA sequencing and subsequent molecular phylogenetic analyses. The fungal DNA barcoding, which uses a standardized 500-800 bp sequence to identify species, has been a popular tool in both biodiversity and phytopathological studies as well as comparatively few cases in plant disease regulatory purposes. Recognition of the ribosomal Internal Transcribe Spacer region (ITS) as the standard DNA barcode for fungi, in 2012 is considered as a milestone in fungal studies with a greater scientific importance (Schoch et al. 2012). However, it should be noted that fungal species identification based on DNA sequence information from a single barcode locus, deviates from the current gold standard for fungal species recognition that utilizes the phylogenetic approach and is based on the concordance of multiple gene genealogies (Taylor et al. 2000). Today's molecular identification of fungi has evolved from DNA barcoding to DNA-aided taxonomy and phylogeny based on the premise of using the genetic variation inherent among sequences of different individuals to identify taxa (Figure 2).

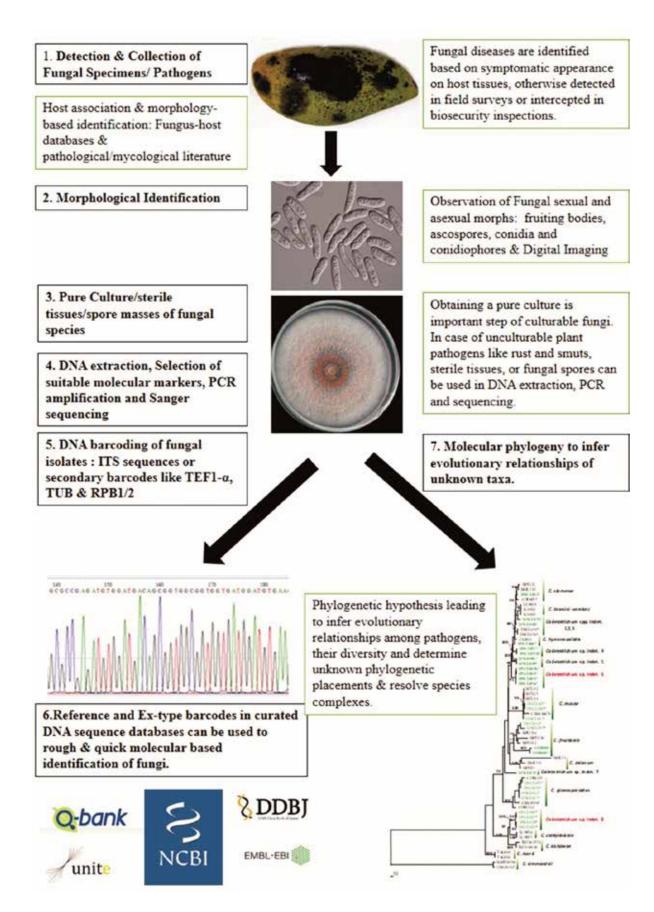


Figure 2: Diagrammatic illustration of the key steps of molecular identification of the plant pathogenic fungi.

The detection, morphological identification and generation of molecular data required skills and involvement of plant pathologists or mycologists. DNA barcodes generated should be carefully handled in order to maintain the quality standards. The International Nucleotide Sequence Database Collaboration (INSDC) is a long-standing foundational initiative that operates between DDBJ, EMBL-EBI and NCBI available for sequence similarity search. The Q-bank Fungi database contains curated DNA sequence data (barcodes), morphological, phenotypical and ecological data for approximately 600 species that are relevant to mycological phytopathology. The UNITE is a wellcurated nucleotide database supported with ITS based Species Hypotheses (SHs), with collection of annotated ITS sequences. Phylogenetic reconstructions can be used to place unknown taxa within current classification system and predict relationships among them.

Phylogenetic species recognition of an unknown is established based on evolutionary hypothesis, thus advocating a robust biological basis for recognizing fungal species. For instance, the boxwood blight, caused by the ascomycetous fungus, Calonectria pseudonaviculata, is a recently known emerging disease, particularly in the USA infecting ornamental and native boxwood plants in the family Buxaceae (Ivors et al. 2012; Henricot & Wedgwood 2013). The disease has been first reported from a single location at a single nursery in southern England in 1994, and it is now known throughout Europe, Asia, New Zealand, and North America (Henricot & Culham 2002). The first documented and confirmed reports of the disease from USA were made from Connecticut and North Carolina in November 2011, followed by reporting in more than 10 other states and on alternative hosts within the United States (LeBlanc et al. 2018; Malapi-Wight et al. 2014). This disease has concerned the ornamental plant industry, due to severe threats to health and productivity of boxwood in both landscape plantings and nurseries (Gehesquière et al. 2016; LeBlanc et al. 2018). The studies on routine collections, molecular and morphological characterization as well as molecular markers aided studies of population structure have proven a substantial impact on the epidemiology and control of this destructive disease within United States and elsewhere in the world (Gehesquière et al. 2016).

Species of *Colletotrichum* are commonly encountered fungal pathogens associated with fruits

and a wide range of field crops and produce anthracnose symptoms and leaf spots, rots and seedling infections (Hyde et al. 2009; Cannon et al. 2012). Since the recommendations of polyphasic approach to study fungal pathogens, coupled with multi-locus phylogenetic reconstructions, the number of phylogenetic species within this genus has rapidly grown (Cai et al. 2009). Many species of Colletotrichum previously considered to be a single taxon have turned out to be "species complexes" consisting of multiple taxa. For instance, Colletotrichum acutatum is generally accepted to cause anthracnose of strawberries (Fragaria × ananassa) (Baroncelli et al. 2015; Damm et al. 2012). Damm et al. (2012), who performed a comprehensive molecular phylogenetic study, revealed that several species in the C. acutatum complex namely, C. acutataum, C. fioriniae, C. nymphaeae, C. simmondsii, C. godetiae and C. salicis, are associated with strawberry diseases. The anthracnose of banana is known to cause by Colletotrichum musae, which is commonly reported from tropical regions of the world where Musa species are widely grown (Ploetz, 1997). Multi-gene phylogenetic analysis revealed that C. musae groups in a distinct lineage within the Colletotrichum gloeosporioides species complex and is most closely related to C. fructicola (Su et al. 2011; Udayanga et al. 2013). This particular species exclusively found on banana species, and considered as a relatively host-specific pathogen, which is an aggressive, difficult to control postharvest disease (Privadarshanie & Vengadaramana 2015). However, the recent studies of Colletotrichum species associated with banana particularly in tropics revealed that several species can be found associated with anthracnose, which make it more crucial in identifying, regulation and control (Vieira et al. 2017). For instance, latest reports of Colletotrichum species associated with banana anthracnose in Brazil confirmed the encounter of C. chrysophilum, C. tropicale, C. theobromicola, and C. siamense in association with the same disease along with C. musae (Vieira et al. 2017).

Fusarium oxysporum, is one of the most economically important and commonly encountered fungal species associated with plants, animals and human as both pathogenic and non-pathogenic strains (Al-Hatmi et al. 2015; Nirmaladevi et al. 2017; Gordon 2017). This species was originally described from Germany, causing dry rot of *Solanum tuberosum* (potato) (Von Schlechtendahl 1824). Subsequently, the use of sub-specific ranks (special forms or formae speciales) in F. oxysporum has been popular practice among plant pathologists due to the broad morphological diversity and host range (Leslie & Summerell 2006). Some of the formae specialis of F. oxysporum cause noteworthy diseases on economically important crops worldwide. For instance, global banana production is seriously threatened by the Fusarium Wilt disease, caused by the soil-borne pathogen Fusarium oxysporum f. sp. cubense (Foc) and also known as "Panama disease" (Mostert et al. 2017). This disease wiped out the banana industry in Central America and the Caribbean, in the mid-twentieth century and is now reported from many of the banana growing tropical countries across the globe (De La Cruz & Jansen 2018). The previous system of special forms of F. oxysporum, has much confused the identification of this important fungal pathogen due to subjectivity of morphological identification and intricate genetic variations. Recently, considering the need to advance and stabilize the taxonomic position of F. oxysporum, an epitype originally collected from Germany from Solanum was designated for this species (Lombard et al. 2019). In the same study, a combined four gene phylogeny based on the loci cmdA (calmodulin), rpb2 (DNAdirected RNA polymerase II), tef1 (translation elongation factor -1) and tub2 (tubulin-2) genes revealed that the isolates form Musa classified as f. sp. cubense belong in at least four different clades corresponding to distinct species, Fusarium duoseptatum, F. nirenbergiae, F. odoratissimum and F. trachichlamydosporum (Lombard et al. 2019). The designation of an epitype, supported by substantial molecular phylogenetic evidence, allows naming of the multiple cryptic species recognized in this species complex, thus plant pathologists as well as biosecurity officials can adopt robust standards in their decisions regarding regulations of pathogens (Wingfield et al. 2012).

Need for Paradigm Changes in Global Plant Biosecurity Practices

International trade of horticultural products and living plants has allowed the movement of plant associated fungi across the borders for centuries. The quick identification of an invasive species is a first line of defense; however, when it comes to fungi, accurate identification is often problematic with minimal macro-morphological features or microscopic observations. Many of the checklists and databases of invasive and quarantine fungi as well as disease compendia have become outdated due to the identification of species merely based on morphology. To ensure the plant biosecurity of the country where sensitive cases of accurate fungal identification are vital, robust consolidated morphological and molecular based identification protocols should be followed (Aveskamp et al. 2008). This will not only assure the health of crops and natural ecosystems but also prevent unnecessary trade regulations for export-agriculture based economies. Almost all molecular plant pathology and phylogenetic studies of fungi frequently emphasize the significance of downstream utilization of their research outcome to ensure the sustainable plant health and global food security (Crous et al. 2016; Marsberg et al. 2017). However, much of the information available from research and thousands of DNA barcode sequences available in nucleotide databases may not be actually exploited for biosecurity purposes in many countries especially in highly vulnerable regions for severe food security issues. Although DNA sequencing technologies have become more accessible and affordable for the scientific community, the implementation of these golden tools in plant biosecurity, disease surveillance and quarantine are largely overlooked.

Apart from the dogma of taxon based identification and phylogenetics of plant pathogenic fungi, recent developments in research focused on genomes, transcriptomes, proteomes, and metabolomes of fungi in conjunction with biological information, can be used to determine whether a specific group of fungi poses a biosecurity risk (McTaggart et al. 2016). Fungal genomics can be used to predict the life modes of poorly known fungi, potential pathogenicity factors, including host target proteins and the presence of transposable elements with high mutation rates (McTaggart et al. 2016; Udayanga et al. 2017). Therefore, the biosecurity related decisions only based minimum morphological observations and tentative identifications, may largely underestimate the potential risks of high priority pathogens as well as other emerging species. Topics on biosecurity, plant disease surveillance and quarantine can be involved in a sequential approach of the awareness of the problem, involvement among plant pathologists and other experts, strong collaborative efforts and support from government agencies, the commercial sector, international organizations, research institutes and universities. We expect this article will stimulate further interest, research and discussion of this topic within scientific

communities, policy makers, other relevant authorities and the general public.

Acknowledgement

The author is grateful to Dr. Lisa A. Castlebury (The Agricultural Research Service-United States Department of Agriculture, Beltsville, Maryland, USA) for constructive criticisms of the manuscript.

References

Al-Hatmi, AM, Hagen, F, Menken, SB, Meis, JF & De Hoog, GS 2016. Global molecular epidemiology and genetic diversity of *Fusarium*, a significant emerging group of human opportunists from 1958 to 2015, *Emerging microbes & infections*, vol. 5, no.1, pp. 1–11.

Avelino, J, Cristancho, M, Georgiou, S, Imbach, P, Aguilar, L, Bornemann, G, Läderach, P, Anzueto, F, Hruska, AJ & Morales, C 2015. The coffee rust crises in Colombia and Central America (2008– 2013): impacts, plausible causes and proposed solutions, *Food Security*, vol. 7, no.2, pp. 303–321. Aveskamp, MM, De Gruyter, J & Crous, PW 2008.Biology and recent developments in the systematics of Phoma, a complex genus of major quarantine significance, *Fungal Diversity*, vol. 31, pp. 1–18.

Baroncelli, R, Sarrocco, S, Zapparata, A, Tavarini, S, Angelini, LG & Vannacci, G 2015. Characterization epidemiology of and Colletotrichum acutatumsensu (C.lato chrysanthemi) causing Carthamus tinctorius anthracnose, Plant pathology, vol. 64, no. 2, pp. 375-384.

Beenken, L 2017. *Austropuccinia*: a new genus name for the myrtle rust *Puccinia psidii* placed within the redefined family *Sphaerophragmiaceae* (Pucciniales), *Phytotaxa*, vol. 297 no. 1, pp. 53–61. Burgess, TI, Crous, CJ, Slippers, B, Hantula, J & Wingfield, MJ 2016. Tree invasions and biosecurity: eco-evolutionary dynamics of hitchhiking fungi, *AoB Plants*, vol. 8.

Cai, L, Hyde, KD, Taylor, PWJ, Weir, B, Waller, J, Abang, MM, Zhang, JZ, Yang, YL, Phoulivong, S, Liu, ZY & Prihastuti, H 2009. A polyphasic approach for studying Collectotrichum, *Fungal Diversity*, vol. *39*, no. 1, pp. 183–204.

Cai, L, Giraud, T, Zhang, N, Begerow, D, Cai, G & Shivas, RG 2011. The evolution of species concepts and species recognition criteria in plant pathogenic fungi, *Fungal Diversity*, vol. *50*, no. 1, pp. 121–133.

Cannon, PF, Damm, U, Johnston, PR & Weir, BS 2012. *Colletotrichum*–current status and future directions, *Studies in mycology*, vol. 73, pp.181–213.

Carvalho, GMA, Carvalho, CR, Barreto, RW & Evans, HC 2014. Coffee rust genome measured using flow cytometry: does size matter?. *Plant pathology*, vol. *63*, no.5, pp. 1022–1026.

Couch, BC & Kohn, LM 2002. A multilocus gene genealogy concordant with host preference indicates segregation of a new species, *Magnaporthe oryzae*, from *M. grisea*. *Mycologia* vol. 94: pp. 683–693.

Cressey, D 2013. Coffee rust regains foothold, *Nature News*, vol. *493*, no. 7434, pp. 587.

Crous, PW, Hawksworth, DL & Wingfield, MJ 2015. Identifying and naming plant-pathogenic fungi: past, present, and future, *Annual Review of Phytopathology*, vol. *53*, pp. 247–267.

Crous, PW, Groenewald, JZ, Slippers, B & Wingfield, MJ 2016. Global food and fibre security threatened by current inefficiencies in fungal identification, *Philosophical Transactions of the Royal Society B: Biological Sciences*, vol. *371*, no. 1709, p.20160024.

Damm, U, Cannon, PF, Woudenberg, JHC & Crous, PW 2012. The *Colletotrichum acutatum* species complex, *Studies in mycology*, vol. *73*, pp.37–113.

De La Cruz, J & Jansen, K 2018, Panama disease and contract farming in the Philippines: Towards a political ecology of risk, *Journal of Agrarian Change*, vol. *18*, *no*. 2, pp. 249–266.

Dean, R, Van Kan, JA, Pretorius, ZA, Hammond-Kosack, KE, Di Pietro, A, Spanu, PD, Rudd, JJ, Dickman, M, Kahmann, R, Ellis, J & Foster, GD 2012, The Top 10 fungal pathogens in molecular plant pathology, *Molecular plant pathology*, vol. *13*, *no.* 4, pp. 414–430.

Doehlemann, G, Ökmen, B, Zhu, W & Sharon, A 2017, Plant Pathogenic Fungi. *Microbiology spectrum*, vol. *5, no.* 1.

Ehrenbergerová, L, Kučera, A, Cienciala, E, Trochta, J & Volařík, D 2018, Identifying key factors affecting coffee leaf rust incidence in agroforestry plantations in Peru. *Agroforestry systems*, vol. *92, no.* 6, pp. 1551–1565.

Elad, Y & Pertot, I 2014, Climate change impacts on plant pathogens and plant diseases. *Journal of Crop Improvement*, vol. 28, no. 1 pp. 99139.

Gehesquière, B, Crouch, JA, Marra, RE, Van Poucke, K, Rys, F, Maes, M, Gobin, B, Höfte, M & Heungens, K 2016, Characterization and taxonomic reassessment of the box blight pathogen *Calonectria pseudonaviculata*, introducing *Calonectria* *henricotiae* sp. nov. *Plant Pathology*, vol. 65, no. 1, pp. 37–52.

Geiser, DM, Aoki, T, Bacon, CW, Baker, SE, Bhattacharyya, MK, Brandt, ME, Brown, DW, Burgess, LW, Chulze, S, Coleman, JJ & Correll, JC 2013, One fungus, one name: defining the genus Fusarium in a scientifically robust way that preserves longstanding use. *Phytopathology*, vol. *103, no.* 5, pp. 400–408.

Godfray, HCJ, Mason-D'Croz, D & Robinson, S 2016, Food system consequences of a fungal disease epidemic in a major crop.*Phil. Trans. R. Soc. B*, vol. 371, no. 1709, p.20150467.

Gordon, TR 2017, *Fusarium oxysporum* and the *Fusarium* wilt syndrome. *Annual review of phytopathology*, vol. 55, pp. 23–39.

Hawksworth, DL 2015, Naming fungi involved in spoilage of food, drink, and water. *Current Opinion in Food Science*, vol. *5*, pp. 23–28.

Hawksworth, DL 2011, A new dawn for the naming of fungi: impacts of decisions made in Melbourne in July 2011 on the future publication and regulation of fungal names. *IMA fungus: the global mycological journal*, vol. *2*, *no.* 2, p. 155.

Hawksworth, DL, Crous, PW, Redhead, SA, Reynolds, DR, Samson, RA, Seifert, KA, Taylor, JW & Wingfield, MJ 2011, The Amsterdam declaration on fungal nomenclature. *IMA fungus*, vol. 2, no. 1, pp. 105–112.

Henricot, B & Culham, A 2002, *Cylindrocladium buxicola*, a new species affecting *Buxus* spp., and its phylogenetic status. *Mycologia*, vol. 94, pp. 980–997.

Henricot, B & Wedgwood, E, 2013, Evaluation of foliar fungicide sprays for the control of boxwood blight, caused by the fungus *Cylindrocladium buxicola*. *Plant Health Progress*, vol. *14*, no. 1, p. 2. Hibbett, DS, Binder, M, Bischoff, JF, Blackwell, M, Cannon, PF, Eriksson, OE & Lumbsch, HT 2007, A higher-level phylogenetic classification of the Fungi. *Mycological research*, vol. *111*, no. 5, pp. 509–547.

Hibbett, D, Abarenkov, K, Kõljalg, U, Öpik, M, Chai, B, Cole, J, Wang, Q, Crous, P, Robert, V, Helgason, T & Herr, JR 2016, Sequence-based classification and identification of Fungi. *Mycologia*, vol. *108, no.* 6, pp. 1049–1068.

Hyde, KD, Cai, L, McKenzie, EHC, Yang, YL, Zhang, JZ & Prihastuti, H 2009, *Colletotrichum*: a catalogue of confusion. *Fungal Diversity*, vol. *39*, no. 1.

Hyde, KD, Udayanga, D, Manamgoda, DS, Tedersoo, L, Larsson, E, Abarenkov, K, Bertrand, YJ, Oxelman, B, Hartmann, M, Kauserud, H & Ryberg, M 2013, Incorporating molecular data in fungal systematics: a guide for aspiring researchers. vol. *3*, no. 1, pp. 11-32.

Ivors, KL, Lacey, LW, Milks, DC, Douglas, SM, Inman, MK, Marra, RE & LaMondia, JA 2012, First report of boxwood blight caused by Cylindrocladium pseudonaviculatum in the United States. Plant Disease, vol. 96, no. 7, pp. 1070-1070. Khan, MAI, Ali, MA, Monsur, MA, Kawasaki-Tanaka, A, Hayashi, N, Yanagihara, S, Obara, M, Mia, MAT, Latif, MA & Fukuta, Y 2016, Diversity and distribution of rice blast (Pyricularia oryzae Cavara) races in Bangladesh. Plant disease, vol. 100 no. 10, pp. 2025-2033.

LeBlanc, N, Salgado-Salazar, C & Crouch, JA 2018, Boxwood blight: an ongoing threat to ornamental and native boxwood. *Applied microbiology and biotechnology*, vol. *102*, *no*. 10, pp. 4371-4380.

Leslie, JF & Summerell, BA 2006, Fusarium laboratory workshops—A recent history. *Mycotoxin Research*, vol. 22, no. 2, pp. 73–74.

Lombard, L, Sandoval-Denis, M, Lamprecht, SC & Crous, PW 2019, Epitypification of Fusarium oxysporum–clearing the taxonomic chaos. *Persoonia-Molecular Phylogeny and Evolution of Fungi*, vol. 43, pp. 1–47.

Malapi-Wight, M, Hébert, JB, Buckley, R, Daughtrey, ML, Gregory, NF, Rane, K, Tirpak, S & Crouch, JA 2014, First report of boxwood blight caused by *Calonectria pseudonaviculata* in Delaware, Maryland, New Jersey and New York. *Plant Dis*, vol. *98*, p. 698.

Marsberg, A, Kemler, M, Jami, F, Nagel, JH, Postma-Smidt, A, Naidoo, S, Wingfield, MJ, Crous, PW, Spatafora, JW, Hesse, CN & Robbertse, B 2017, *Botryosphaeria dothidea*: a latent pathogen of global importance to woody plant health. *Molecular plant pathology*, vol. *18*, no. 4, pp. 477–488.

McCook, S & Vandermeer, J 2015, The big rust and the red queen: long-term perspectives on coffee rust research. *Phytopathology*, vol. *105, no.* 9, pp. 1164–1173.

McCook, S 2006, Global rust belt: *Hemileia vastatrix* and the ecological integration of world coffee production since 1850. *Journal of Global History*, vol. *1*, no. 2, pp.177–195.

McDonald, BA & Stukenbrock, EH 2016, Rapid emergence of pathogens in agro-ecosystems: global threats to agricultural sustainability and food security. *Philosophical Transactions of the Royal Society B: Biological Sciences*, vol. 371, no. 1709, p.20160026. McTaggart, AR, van der Nest, MA, Steenkamp, ET, Roux, J, Slippers, B & Shuey, LS 2016, Fungal Genomics Challenges the Dogma of Name-Based Biosecurity. *PLoS Pathog* vol. 12, no. 5: e1005475. Morin, L, Aveyard, R, Lidbetter, JR & Wilson, PG 2012, Investigating the host-range of the rust fungus *Puccinia psidii sensu lato* across tribes of the family Myrtaceae present in Australia. *PLoS One*, vol. 7, no. 4, p.e35434.

Mostert, D, Molina, AB, Daniells, J, Fourie, G, Hermanto, C, Chao, CP, Fabregar, E, Sinohin, VG, Masdek, N, Thangavelu, R & Li, C 2017, The distribution and host range of the banana Fusarium wilt fungus, *Fusarium oxysporum* f. sp. *cubense*, in Asia. *PloS one*, vol. *12, no.* 7, p.e0181630.

Nalley, L, Tsiboe, F, Durand-Morat, A, Shew, A & Thoma, G 2016, Economic and environmental impact of rice blast pathogen (*Magnaporthe oryzae*) alleviation in the United States.*PloS one*, vol. 11, no. 12, p.e0167295.

Nirmaladevi, D, Venkataramana, M, Srivastava, RK, Uppalapati, S.R, Gupta, VK, Yli-Mattila, T, Tsui, KC, Srinivas, C, Niranjana, SR & Chandra, NS 2016, Molecular phylogeny, pathogenicity and toxigenicity of *Fusarium oxysporum* f. sp. *lycopersici. Scientific reports*, vol. *6*, p.21367.

Norvell, LL 2011, Fungal nomenclature. 1. Melbourne approves a new Code. *Mycotaxon*, vol. *116*, no. 1, pp.481–490.

Paini, DR, Sheppard, AW, Cook, DC, De Barro, PJ, Worner, SP & Thomas, MB 2016, Global threat to agriculture from invasive species. *Proceedings of the National Academy of Sciences*, vol. *113, no.* 27, pp.7575–7579.

Park, RF, Keane, PJ, Wingfield, MJ, & Crous, PW 2000, Fungal diseases of eucalypt foliage. *Diseases and pathogens of eucalypts*, pp. 153–239.

Ploetz, R 1997, November. Banana diseases in the subtropics: a review of their importance, distribution and management. In *II International Symposium on Banana: I International Symposium on Banana in the Subtropics vol. 490*, pp. 263–276.

Priyadarshanie, HR & Vengadaramana, A 2015, Some preliminary studies of *Colletotrichum musae* associated with banana anthracnose disease in Jaffna district, Sri Lanka. *Universal Journal of Agricultural Research*, vol. *3*, no. 6, pp. 197-202.

Raja, HA, Miller, AN, Pearce, CJ & Oberlies, NH 2017, Fungal identification using molecular tools: a primer for the natural products research community. *Journal of natural products*, vol. *8*, no. 3, pp.756–770.

Rossman, AY & Palm-Hernández, ME 2008, Systematics of plant pathogenic fungi: why it matters. *Plant Disease*, vol. *92, no.* 10, pp. 1376–1386.

Rossman, AY, Farr, DF & Castlebury, LA 2007, A review of the phylogeny and biology of the Diaporthales. *Mycoscience*, vol. 48, no. 3, pp. 135–144.

Rossman, AY, Allen, WC & Castlebury, LA 2016, New combinations of plant-associated fungi resulting from the change to one name for fungi. *IMA fungus*, vol. 7, no. 1, pp. 1–7.

Saleh, D, Milazzo, J, Adreit, H, Fournier, E & Tharreau, D 2014, South-East Asia is the center of origin, diversity and dispersion of the rice blast fungus, *Magnaporthe oryzae*. *New Phytologist*, vol. *201*, no. 4, pp. 1440–1456.

Savary, S, Willocquet, L, Elazegui, FA, Castilla, NP & Teng, PS 2000, Rice pest constraints in tropical Asia: quantification of yield losses due to rice pests in a range of production situations. *Plant disease*, vol. *84*, no. 3, pp.357–369.

Schoch, CL, Seifert, KA, Huhndorf, S, Robert, V, Spouge, JL, Levesque, CA, Chen, W & Fungal Barcoding Consortium, 2012, Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences*, vol. *109*, no. 16, pp. 6241–6246.

Stricker, KB, Harmon, PF, Goss, EM, Clay, K & Luke Flory, S 2016, Emergence and accumulation of novel pathogens suppress an invasive species. *Ecology Letters*, vol. 19, no.4, pp. 469–477. Su, YY, Noireung, P, Liu, F, Hyde, KD, Moslem, MA, Bahkali, AH, Abd-Elsalam, KA & Cai, L 2011, Epitypification of Colletotrichum musae, the causative agent of banana anthracnose. *Mycoscience*, vol. 52, no. 6, pp. 376– 382.

Taylor, JW, Jacobson, DJ, Kroken, S, Kasuga, T, Geiser, DM, Hibbett, DS & Fisher, MC 2000, Phylogenetic species recognition and species concepts in fungi. *Fungal genetics and biology*, vol. *31*, no.1, pp. 21–32.

Taylor, JW 2011, One fungus one name: DNA and fungal nomenclature twenty years after PCR. *IMA fungus*, vol. 2, no. 2, pp. 113–120.

Tedersoo, L, Bahram, M, Põlme, S, Kõljalg, U, Yorou, NS, Wijesundera, R, Ruiz, LV, Vasco-Palacios, AM, Thu, PQ, Suija, A & Smith, ME 2014, Global diversity and geography of soil fungi. *Science*, vol. 346, no. 6213, p.1256688.

Udayanga, D, Liu, X, McKenzie, EH, Chukeatirote, E, Bahkali, AH & Hyde, KD 2011, The genus

Phomopsis: biology, applications, species concepts and names of common phytopathogens. *Fungal diversity*, vol. *50, no.* 1, pp. 189–225.

Udayanga, D, Manamgoda, DS, Liu, X, Chukeatirote, E & Hyde, KD 2013, What are the common anthracnose pathogens of tropical fruits?. *Fungal Diversity*, vol. *61, no.* 1, pp. 165–179.

Udayanga, D, Castlebury, LA, Rossman, AY & Hyde, KD 2014, Species limits in Diaporthe: molecular re-assessment of *D. citri*, *D. cytosporella*, *D. foeniculina* and *D. rudis*. *Persoonia: Molecular Phylogeny and Evolution of Fungi*, vol. *32*, pp.83.

Udayanga, D & Castlebury, LA 2017, Molecular systematics and comparative draft genome analysis of strawberry leaf blight fungus. *International Research Symposium on Pure and Applied Sciences*, 2017 Faculty of Science, University of Kelaniya, Sri Lanka. pp.15.

Vandermeer, J, Hajian-Forooshani, Z & Perfecto, I 2018, The dynamics of the coffee rust disease: an epidemiological approach using network theory. *European journal of plant pathology*, vol. *150*, no. 4, pp.1001–1010.

Vandermeer, J, Perfecto, I & Philpott, S 2010, Ecological complexity and pest control in organic coffee production: uncovering an autonomous ecosystem service. *BioScience*, vol. 60, no. 7, pp. 527–537.

Vieira, WADS, Lima, WG, Nascimento, ES, Michereff, SJ, Reis, A, Doyle, VP & Câmara, MPS 2017, Thiophanate-methyl resistance and fitness components *of Colletotrichum musae* isolates from banana in Brazil. Plant disease, vol. 101, no. 9, pp. 1659–1665.

Von Schlechtendahl FK 1824. Flora berolinensis.Pars secunda. *Cryptogamia*, Berlin. vol. 2, pp. 139.

Wingfield, MJ, De Beer, ZW, Slippers, B, Wingfield, BD, Groenewald, JZ, Lombard, L & Crous, PW 2012, One fungus, one name promotes progressive plant pathology. *Molecular plant pathology*, vol. *13*, no. 6, pp. 604–613.